

ECOLOY OF PALYTHOA CAESIA (ANTHOZOA: ZOANTHIDEA) IN TURBID ENVIRONMENT

NAPALAI JUNTARUK

A THESIS PRESENTED TO RAMKHAMHAENG UNIVERSITY
IN PARTIAL FULFILLMENT OF THE REQUIREMENTS
FOR THE DEGREE OF MASTER OF SCIENCE
(BIOLOGY)
2003

1 0 W.A. 2547



โครงการพัฒนาองต์ความรู้และศึกษานโยบายการจัดการทรัพยากรชิวภาพในประเทศไทย c/o ดูนย์พันธุวิตวกรรมและเทคโนโลยีชีวภาพแห่งชาติ อาการสำนักงานพัฒนาวิทยาศาสตร์และเทคโนโลยีแห่งชาติ 73/1 ถนนพระรามที่ 6 เขตราชเทวี กรุงเทพฯ 10400

ECOLOY OF *PALYTHOA CAESIA* (ANTHOZOA: ZOANTHIDEA) IN TURBID ENVIRONMENT

NAPALAI JUNTARUK

A THESIS PRESENTED TO RAMKHAMHAENG UNIVERSITY IN PARTIAL FULFILLMENT OF THE REQUIREMENTS FOR THE DEGREE OF MASTER OF SCIENCE (BIOLOGY)

2003

CORYRIGHTED BY RAMKHAMHAENG UNIVERSITY
ISBN 974-09-1077-7

นิเวศวิทยาของพรมทะเล *Palythoa ceasia* (Anthozoa: Zoanthidea) ในสภาพแวคล้อมที่มีความขุ่น

นภาลัย จันทรักษ์

วิทยานิพนธ์เสนอต่อมหาวิทยาลัยรามคำแหง
เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญา
วิทยาศาสตรมหาบัณฑิต (ชีววิทยา)
ปีการศึกษา 2546
ลิขสิทธิ์ของมหาวิทยาลัยรามคำแหง
ISBN 974-09-1077-7

Thesis Title	Ecology of Palythoa ceasia (Anthozoa:
	Zoanthidea) in Turbid Environment
Student's Name	Miss Napalai Juntaruk
	Department of Biology, Faculty of Science
Advisory Committee	Dr. Thamasak Yeemin Chairperson
	Assoc. Prof. Supawadee Chullasorn
	Asst. Prof. Dr. Sutat Subinprasert
Ramkhamhaeng Ur	niversity approved this thesis as a partial fulfillment
of the requirements for the	e degree of Master of Science (Biology)
	Dean of Graduate School
	Suraphol Rajbhandaraks)
Examination Committee	
Tha	mand Ye Chairperson
	amasak Yeemin)
5.	Member
(Assoc. Prof	Supawadee Chullasorn)
Jeu	tet Labinot Member
(Asst. Prof.	Dr. Sutat Subinprasert)

1344010

ABSTRACT

Thesis Title

: Ecology of Palythoa ceasia (Anthozoa :

Zoanthidea) in Turbid Environment

Student's Name

: Miss Napalai Juntaruk

Degree Sought

: Master of Science

Major

: Biology

Academic Year

: 2003

Advisory Committee:

1. Dr. Thamasak Yeemin

Chairperson

2. Assoc. Prof. Supawadee Chullasorn

3. Asst. Prof. Dr. Sutat Subinprasert

Ecology of *Palythoa ceasia* (Anthozoa: Zoanthidea) in turbid environment at Khang Khao Island, Chonburi Province was studied during June 2000 – February 2002. *P. ceasia* plays a major role as a dominant competitor with sessile reef organisms. It can overgrowth other sessile reef invertebrates because of its rapid growth and propagation by both sexual and asexual reproductions. *P. ceasia* can assimilate sediment on colony surface and in its mesoglea. This study concentrated on distribution patterns, reproductive ecology, growth and mortality rates, sediment assimilation and competition ability of *P. caesia*. The results showed that mean growth rate sediment

assimilation and competition ability of P. caesia was 1.45 cm./mo. While mean mortality rate was 25.08% per year. Change of coverage during June 2000 - February 2002 was + 0.41% per year. P. ceasia was a hermaphrodite. Average of sedimentation on P. ceasia colony surface in deep stations was $3,442.85 \pm 2,173.60$ mg./m. while that of shallow stations was $2,931.81 \pm$ 2,787.49 mg./m.². Average of assimilated sediment in deep stations was $15,881.60 \pm 5,837.43$ mg./m. but that of shallow stations was $14,564.62 \pm$ 6,777.38 mg./m.² Average sediment found in mesoglea was 71.85% of tissue dry weight. Average of sedimentation rates in the filed measured by using sediment traps with 1.5 cm. and 5 cm. in diameters in deep stations were 58.91 ± 16.41 (mean \pm SE.) and 113.22 ± 21.01 (mean \pm SE.) mg./cm.²/day, respectively, while those in shallow stations were 58.35 ± 13.91 (mean + SE.) and 80.85 ± 18.91 (mean + SE.) mg./cm.²/day, respectively. Sedimentation rates measured by using sediment plates were very low compared to sediment traps. The present research highlighted the importance of P. caesia in coral community changes in the Gulf of Thailand.

7-344010

บทคัดย่อ

ชื่อเรื่องวิทยานิพนธ์ : นิเวศวิทยาของพรมทะเล Palythoa ceasia

(Anthozoa : Zoanthidea) ในสภาพแวคล้อมที่มีความขุ่น

ชื่อผู้เขียน

: นางสาวนภาลัย จันทรักษ์

ชื่อปริญญา

: วิทยาศาสตรมหาบัณฑิต

สาขาวิชา

• ชีววิทยา

ปีการศึกษา

. 2546

คณะกรรมการที่ปรึกษาวิทยานิพนซ์

1. คร.ธรรมศักดิ์ ยีมิน

ประชานกรรมการ

2. รองศาสตราจารย์สุภาวคี จุลละศร

3. ผู้ช่วยศาสตราจารย์ คร.สุทัศน์ สุบินประเสริฐ

การศึกษานิเวศวิทยาของพรมทะเล Palythoa ceasia บริเวณเกาะค้างคาว จังหวัด ชลบุรี ซึ่งเป็นบริเวณที่มีความขุ่นสูง โดยทำการศึกษาตั้งแต่เคือนมิถุนายน 2543 – กุมภาพันธ์ 2545 พรมทะเลมีบทบาทสำคัญในความสัมพันธ์แบบแก่งแย่งในแนว ปะการัง ซึ่งสามารถเจริญเติบโตขึ้นคลุมทับสิ่งมีชีวิตอื่นได้ดี เนื่องจากเจริญเติบโตได้ อย่างรวคเร็ว และสามารถสืบพันธุ์ได้ทั้งแบบอาศัยเพศและไม่อาศัยเพศ พรมทะเลมี ความสามารถในการกักเก็บตะกอนบนผิวของโคโลนีและในชั้น mesoglea การศึกษานี้ มุ่งเน้นศึกษารูปแบบการแพร่กระจาย นิเวศวิทยาการสืบพันธุ์ อัตราการเจริญเติบโต อัตราการตาย ความสามารถในการกักเก็บตะกอน และการแก่งแย่ง ผลการศึกษาพบว่า P. ceasia มีอัตราการเจริญเติบโตเฉลี่ย 1.45 ซม./เคือน อัตราการตายเฉลี่ย 25.08% ต่อปี การเปลี่ยนแปลงพื้นที่การปกคลุมตั้งแต่เคือนมิถุนายน 2543 – กุมภาพันธ์ 2545 เพิ่มขึ้น 0.41% ต่อปี P. ceasia เป็นกะเทย (hermaphrodite) การศึกษาปริมาณตะกอนที่พบบนผิว

ของ P. ceasia บริเวณที่ลึกเฉลี่ย 3,442.85 ± 2,173.60 มก./ต.ร.ม. ที่ตื้นเฉลี่ย 2,931.81 ± 2,787.49 มก./ตร.ม. ปริมาณตะกอนที่สะสมในเนื้อเยื้อบริเวณที่ลึกเฉลี่ย 15,881.60 ± 5,837.43 มก./ตร.ม. ที่ตื้นเฉลี่ย 14,564.62 ± 6,777.38 มก./ตร.ม. ปริมาณตะกอนที่ สะสมในเนื้อเยื่อเฉลี่ยร้อยละ 71.85 ของน้ำหนักแห้ง การศึกษาอัตราการตกตะกอนใน ธรรมชาติจากกับคักตะกอน ขนาดเส้นผ่าศูนย์กลาง 1.5 ซม. บริเวณที่ลึกเฉลี่ย 58.91 ± 9.43 (mean ± SE.) มก./ตร.ซม./วัน บริเวณที่ตื้นเฉลี่ย 58.35 ± 13.91 (mean ± SE.) มก./ตร.ซม./วัน อัตราการตกตะกอนเฉลี่ยจากกับคักตะกอนขนาดเส้นผ่าศูนย์กลาง 5 ซม. บริเวณที่ลึกเฉลี่ย 113.22 ± 21.01 (mean ± SE.) มก./ตร.ซม./วัน บริเวณที่ตื้น เฉลี่ย 80.85 ± 18.91 (mean ± SE.) มก./ตร.ซม./วัน อัตราการตกตะกอนที่ได้จากกับ คักตะกอนแบบจานมีปริมาณน้อยมากเมื่อเทียบกับอัตราการตกตะกอนที่ได้จากกับคัก ตะกอนทั้งสองขนาด ผลการศึกษานี้ชี้ให้เห็นถึงความสำคัญของพรมทะเล P. caesia ที่มี ต่อการเปลี่ยนแปลงของกลุ่มสิ่งมีชีวิตในแนวปะการังของอ่าวไทย

ACKNOWLEDGEMENTS

I would like to express my deepest gratitude to Dr. Thamasak Yeemin, my advisor, for his encouragement and advice throughout the research work; and to Assoc. Prof. Supawadee Chullasorn and Asst. Prof. Dr. Sutat Subinprasert for their helpful guidance and intensive supervision throughout my study. Their kindness and helpfulness will always be remembered.

I am grateful to Department of Biology, Faculty of Science, Ramkhamhaeng University. It is my pleasure to thank a number of colleagues from the excellent organization team, Marine Biodiversity Research Group: Mr. Nisit Ruengsawang, Mr. Saiprateep Asa, Miss. Wasana Phantaewee, Miss. Parnhathai Nopchinwong, Miss. Rattika Petthongma, Miss. Wichanan Phongsri, Miss. Thanida Pinkaew, Mr. Jamrearn Buaruang and Mr. Chaipichit Saenghaisuk for their assistance in the field and laboratory works. I am indebted to Mr. Anuruk Pisaphak for his editing the map in GIS.

This research was supported by TRF/BIOTEC Special Program for Biodiversity Research and Training Grant, BRT T_3440. Finally, I would like to express my profound gratitude to my beloved parents for their love, intensive supervision and encouragement throughout my life.

Napalai Juntaruk

TABLE OF CONTENTS

	Page
ABSTRACT (ENGLISH)	(iv)
ABSTRACT (THAI)	(vi)
ACKNOWLEDGEMENTS	(viii)
LIST OF TABLES	(xii)
LIST OF FIGURES	(xiv)
ABBREVIATIONS	(xvi)
Chapter	
1 INTRODUCTION	1
Objectives	2
Hypothesis	2
Scope of Research	3
2 LITERATRUE REVIEW	4
Morphology of Class Anthozoa	5
Genetics of Class Anthozoa	5
Reproduction	11
Growth rate	15
Competition	16
Environment effect on Anthozoa behavior	18
3 METERIALS AND METHODS	23
Historical background of the Materials	23
Key to Great Barrier Reef and Torres Strait	24

Chapter	Page
Study site	25
Field Experiments	27
Temperature	27
Salinity	28
Sedimentation	28
P. caesia specimen	28
Laboratory works	29
Sediment trap	29
P. caesia colonies	33
Entomb sediment	34
Histological Study	35
4 RESULTS	36
Environmental Factor	36
Salinity	36
Sea Surface Temperature	37
Sedimentation rate and Resuspension of bottom sediment	38
Growth and Mortality rates, Change of colonies size	
and Percent partial Mortality of P. caesia	47
Sexual reproduction of <i>P. caesia</i>	50
Competition of P. caesia.	55
Distribution Patterns of P. caesia	56
Assimilated and Surface sediment in P. caesia	63

Chapter	Page
5 DISCUSSION	67
Growth and Mortality rates	67
Change of P. caesia colony size	68
Reproduction	69
Population density and Distribution pattern	73
Competition	74
Assimilated and surface sediment in P. caesia tissues	75
APPENDIX	76
BIBLIOGRAPHY	82
VITA	90

LIST OF TABLES

[able	e	Page
1	Diagnostic features for morphological grouping of Zoanthidae	
	and one species of Parazoanthid	7
2	Sedimentation rate in sediment traps 1.5 cm. diameter	
	(mg./cm. ² /day.)	40
3	Sedimentation rate in sediment traps 5 cm. diameter	
	(mg./cm. ² /day.)	41
4	Mean amount of inorganic mater from sediment traps (g./ash)	42
5	Mean amount of organic matter from sediment traps (g./DW.)	43
6	Resuspension of bottom sediment rate in sedimentation plates	
	(mg./cm. ² /day.)	46
7	Percent of aggregation of P. caesia with other species	57
8	Amount of assimilated sediment in P. caesia tissues	
-	(g./Ash./m. ²)	65
9	Amount of surface sediment on P. caesia colony (g./DW./m.2)	66
10	Mode of reproduction, sex, oocyte and sperm diameter and	
	reproduction cycle in the different zoanthid studied to date	71
11	Two factor of ANOVA on Growth and Mortality rates of	
	P caesia	77

Table		Page
12	Two factor of ANOVA on 5 cm. diameter sediment traps and	
	sediment plates	77
13	Two factor of ANOVA on 1.5 cm. diameter sediment traps	
	and sediment plates	78
14	Two factor of ANOVA on surface sediment from P. caesia	
	and resuspension of bottom sediment	79
15	Two factor of ANOVA on 1.5 cm. diameter and 5 cm.	
	diameter sediment trap	80
16	Two factor of ANOVA on 1.5 cm. diameter sediment trap and	
	sediment plate	80
17	Two factor of ANOVA on 5 cm. diameter sediment trap and	
	andiment plate	01

LIST OF FIGURES

Figu	re	Page
1	The organelle of Anthozoa	6
2	Palythoa schematic illustration of asexual reproduction by	
	Fragmentation	13
3	Type of overgrowth interactions Palythoa by growing over	
	subordinate species	19
4	Map of the study area	27
5	Study of sedimentation and resuspension rate by three types of	
	sediment traps	29
6	Washing sediment traps by running water	31
7	Place beaker in room temperature	31
8	Dry sediment from a sediment trap at 105°C, 24 hour	32
9	Weighing.	32
10	Dry sample burn in muffle furnace at 550°C, 5 hours to remove	33
11	Sediment from P. caesia surface on a filter paper	34
12	Salinity	36
13	Sea surface temperature	37
14	Sedimentation rates from sedimentation traps, 1.5 cm. diameter	44
15	Sedimentation rates from sedimentation traps, 5 cm. diameter	45
16	Resuspention of bottom sediment rates from sedimentation plates.	46
17	Growth and mortality rates of P. caesia	48

Figure	Page
18 Growth and mortality rates and change in colony sizes in the filed	
study	49
19 Number of fertilized polyps from each sampling periods	51
20 Change of colonies size in P. caesia	51
21 Oocyte and Spermatozoa on the same gonad	52
22 Position of oocyte and spermatozoa on a mesentery	52
23 Group of oocytes in gonad and nucleus in oocytes	53
24 Oocyte containing zooxanthellae before spawning	53
25 Sperm in a histological section	54
26 Mature spermatocyte found in a sample	54
27 Spermatozoa and sperm tail in a histological section	55
28 Number of sessile colonies in aggression with P. caesia	58
29 Percent of aggregation of P. caesia with other reef organisms	58
30 P. caesia tissues grew on dead coral without contact	59
31 P. caesia tissues overgrew on living coral	60
32 Distribution patterns of <i>P. caesia</i> on station C	62
33 Distribution patterns of <i>P. caesia</i> on station D	63
34 Entomb sediment found in <i>P. caesia</i> tissues	64
35 Amount of assimilated sediment in <i>P. caesia</i> tissues	65
36 Amount of surface sediment on P. caesia colony	66

ABBREVIATIONS

 μ m = micrometer

ANOVA = analysis of variance

cm = centimeter

et al. = et alii; and others

i.e. = id est (that is)

ml = milliliter

mm = millimeter

SCUBA = Self-contained underwater breathing apparatus

SE = Standard error

sp. = Species

CHAPTER 1

INTRODUCTION

Coral reefs are the highest biodiversity in the ocean (Allen and Steen, 1999, 9) and have an important role for marine ecosystem. They are more complex of plants, animals and bacteria that make them be a major marine resource. (Burnett et al. 1994, 153-159; Burnett et al. 1997, 55-6767; Berntson et al. 1999, 423) Hard coral, soft coral and zoanthid are components widespread throughout the tropical Indo-Pacific region (Chulabhorn Research Institute and Thai Royal Navy 1998, 34; Tanner 1994, 793-809; 390-397; 2000, 514-519)

A zoanthid, *Palythoa*, is a dominant competitor of other reef sessile organisms (Suckhnek and Green 1981, 679; Karlson 1988a, 31; Hill 1998, 13). *Palythoa* can rapidly grow over reef sessile invertebrates and produce a highly marine toxin as palytoxin (PTX). Palytoxin has an effect on ion-channel, may kill an animal and make a bare space of reef. (Gleibs et al. 1995, 1531; Leech and Habener 1998, 1066-1072; Gleibs and Mebs 1999, 1525; Wu et al. 2003, 55) In addition to competitive ability *Palythoa* can entomb sediment to their mesoglea. (Haywick and Muller 1997, 39) At the present a little information about biology of *Palythoa* such as of classification, ecology, reproductive biology and sediment retention is available.

Palythoa caesia (Cnidaria: Anthozoa: Zoanthidea) is a dominant species in the Gulf of Thailand and Indo-Pacific region (Chulabhorn Research Institute and Thai Royal Navy 1998, 34) Therefore, their interactions with reef sessile organisms are neces-sary to study in details. Ecological data of *P. caesia* are very important for reef management in the future.

Objectives

The major objectives of this study are as follows:

- 1. To study distribution pattern of *P. caesia* and its competitive ability with other reef sessile organisms.
- 2. To study growth rate, mortality rate and reproductive ecology of *P. caesia*.
- 3. To study sediment assimilated ability of *P. caesia* and to apply the finding as fundamental data for coral reef management.

Hypothesis

A zoanthid, *Palythoa caesia*, can grow in high turbid environment and can entomb sediment in its mesoglea. It play an important role for coral reef development.

Scope of Research

The present study concentrated on ecology of *Palythoa caesia*(Anthozoa: Zoanthidea) in high turbid environment at Khang Khao Island,
Chonburi Province.

CHAPTER 2

LITERATURE REVIEW

Kingdom

: Animal

Class

: Anthozoa

Subclass

: Zoantharia

Order

: Zoanthidea

Family

: Zoanthidae

Genus

: Palythoa

Species

: caesia

Diagnostic feature of Palythoa caesia (Burnett et al. 1997, 57-65)

A zoanthid, *Palythoa* (Anthozoa: Zoanthidea) (Allen and Steen 1999, 9; Burnett et al. 1994, 153-159; Burnett et al. 1997, 55-67; Berntson et al. 1999, 423) is a widespread distribution throughout The Gulf of Thailand and the Indo-Pacific. It plays a major role on competition with reef sessile invertebrates (Karlson 1980, 894-898; Suckhnek and Green 1981, 679; Bastidas and Bone 1996, 543).

Morphology of Class Anthozoa

Members of Anthozoa are polypoid but larger and more complex than polyps in other classes. They have a flattened oral disc bearing tentacles and a slitlike mouth leading to a laterally flattened stomodaeum. (Figure 1) Mesenteries, intucked gastrodermis with mesoglea between, divide up the gastrovascular cavities. The edges of the mesenteries are trilobed filaments. The outer filaments carry cilia and the central one carries chidocytes and cells that secrete enzyme. The mesoglea is thick, fibrous and cellular (Barnes et al. 1988, 63-70) that contains bits of debris and reef elements to support the polyps (Borneman 1998, 3).

Genetics of Class Anthozoa

Taxonomic reviews of class Anthozoa have historically been confound-ded by high level of morphological variation in family zoanthidea (Burnett et al. 1997, 55). Morphology of *Palythoa* spp. and *Parazoanthus* spp. are shown in Table 1 (Berntson et al. 1999, 423-427) *Palythoa* has 3 forms but from they are no different between fixed gene or clear differences in gene frequent-cies among them based on genetic analysis (Burnett et al. 1997, 55-68). By using 18s rDNA sequences to study a relationship within the class Anthozoa, the order Actiniaria, Zoanthidea, Scleractinia, Antipatharia and Ceriantharia, as well as the subclass Octocorallia, constitute monophyletic taxa and order

Antipatharia as a sista group of order Zoanthidea (Berntson et al. 1999, 417-433).

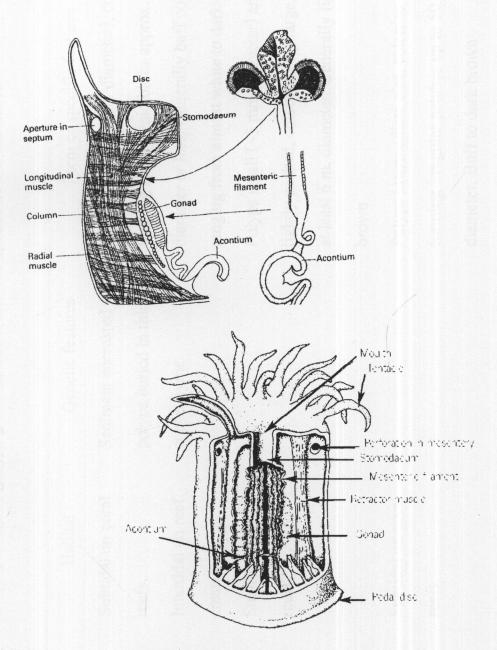


Figure 1 The organelle of Anthozoa

Source: Barnes, R. S. K., P. Calow, P. J. W. Olive, and D. W. Golding.

1988. Phylum Cnidaria (hydras, jellyfish, anemones, corals). In

The Invertebrates. London Blackwell Scientific Press, pp. 55-70.

Table 1 Diagnostic features for morphological grouping of Zoanthidae and one species of Parazoanthid

Morphological group	Habitat	Genetic features	Form or specific features
Palythoa caesia	Subtidal on coral	Sediment encrusted	Polyps large (~8 mm. diameter), colonies
	reefs.	polyps buried in thick	forming small ovoid blobs approx. 10 cm
	Intertidal on reef	coenechyme	diameter or less. Generally buff coloured,
	crests		ranging from almost white to dark brown
Palythoa form 2	Subtidal on coral		Polyps small (~3 mm. diameter) and
	reefs.		tightly packed, colonies very large, up to
			at least 6 m. diameter. Generally light
			brown
Palythoa form 3	Subtidal on coral		Polyps intermediate size, colonies
	reefs.		forming irregular patches up to 50 cm
			diameter. Buff to dark brown

Table 1 (continued)

Morphological group	Habitat	Genetic features	Form or specific features
Protopalythoa mutuki	Intertidal, reef flats	Sediment encrusted	Intertidal. Polyps large, oral disk usually
	and rocky shores	polyps joined only at the	dark green with white mouth. Very
		base.	heavily encrusted with large sand grains
Protopalythoa form 2	Subtidal on coral		Subtidal. Polyps relatively small unless
	reefs.		growing out form under shade. Oral disk
			brown or green, frequently patterned.
Sphenopus marsupialis	Sandy or muddy	Large, solitary polyps	Polyps earthy grey.
	substrata, seagrass	free living in sand. Small	
	beds.	polyps may be attached to	
		stones or sea grass fronds.	

Table 1 (continued)

Morphological group	Habitat	Genetic features	Form or specific features
Zoanthus coppingeri	Intertidal, reef flats	Polyps small, not	Forming extensive colonal patches,
	and rocky shores	sediment encrusted.	especially on unstable substrata (coral
			rubble) and the coral Montipora digitata.
			Colour bright and highly variable,
			generally red, orange, yellow or green
Zoanthus form 2	Intertidal, reef flats		Disk with light coloured pattern on dark
	and rocky shores		green or brown background. Associated
			with Protopalythoa mutuki
Zoanthus form 3	Subtidal on coral		Sutidal. Colonies usually small but
	reefs		occasionally extensive. Disk bright green,
			tentacles brown

Table 1 (continued)

Morphological group	Habitat	Genetic features	Form or specific features
Zoanthus vietnamensis	Intertidal on reef		Colonies thinly encrusting, polyps
	crests		completely buried in coenechyme.
			Forming extensive rubbery mats. Powder
			blue or blue/green.
Parazoanthus dichroicus	Subtidal on coral	Polyps small, encrusted	Growing on hydroids. Polyps small,
	reefs	with fine sediment. Often	beige, with distinct capitular ridges.
		have yellow or orange	Tentacles yellow or orange.
		tentacles.	

Source: Burnett, W. J., J. A. H. Benzie, J. A. Beardmore, and J. S. Ryland. 1997. Zoanthids (Anthozoa,

species. Coral Reefs 16:57.

Hexacorallia) form the Great Barrier Reef and Torres Strait Australia: Systematic, Evolution and key to

Reproduction

Reproduction of the zoanthids can be both sexual and asexual reproductions (Yamazato et al. 1973, 275; Campbell 1974, 133; Cooke 1976, 281; Fadlallah et al. 1984, 80; Martindale and Henry 1998, 672; Borneman and Lowrie 2001, 903). Asexual reproduction of *Palythoa* by using longitudinal budding from stolon (Cooke 1976, 283; Karlson 1988b., 1219; Borneman 1998,3) or fragmentation (Acosta et al. 2001, 363; Borneman and Lowrie 2001, 903) were reported. Acosta et al. (2001, 363-378) examined asexual reproduction of zoanthids and found that zoanthid built a new colonies from fragments. He used *Palythoa caribaeorum* as a model and proposed a way to produce a fragment by two methods.

- 1. Biotic fragmentation (tissue isolated due to biologically causesed lesions); in this case, primary due to disease causing partial colony mortality in the colony (Figure 2 A. C.).
- 2. Physical fragmentation (colonies broken into several pieces by storm, current or tide) (Figure 2 D. E.)

Both methods are incidental fragmentation, not actively directed by the parent colony. A significant positive linear relationship was found between colony area and degree to partial mortality. In general fragmentation in zoanthids plays an important role in asexual reproduction and competition with reef sessile organisms.

In addition to propagation of *P. caesia* by both sexual and asexual modes (Ryland and Babcock 1991, 119). The colonies cannot be identified

their sex. Zoanthid gametes developed in mesentery (Borneman 1998, 3). The eggs and spermaris were different (Cooke 1976, 283). Examples of asexual reproduction were reported in zoanthids.

Yamazato et al. (1973, 276-283) studied reproductive ecology of *Palythoa tuberculosa* in Okinawa, Japan and recognized 3 types of colonies.

Type 1 Functional protogynous hermaphrodites in which the individual polyps change their reproductive phases from functional female to functional hermaphrodite, and then the function male.

Type 2 Consecutive protogynous hermaphrodites in which polyps change from non-functional female to functional male without an intervening period of hermaphroditism.

Type 3 Male colonies, which have only male polyps along with sterile ones.

The above conclusion explains why male colonies are found more frequently than female colonies.

Cooke (1976, 281-288) studied reproductive biology of zoanthids, Zoanthus pacificus and Palythoa vestitus, at Kaneoh Bay, Hawaii and found that the eggs and sperm were very similar in both species. Although fertile polyps for zoanthids were observed throughout the year they did not reveal a seasonal cycle. Gonad developed during summer months and undeveloped in winter months. Both zoanthids had high rates of asexual reproduction by budding new colony near the original colony.

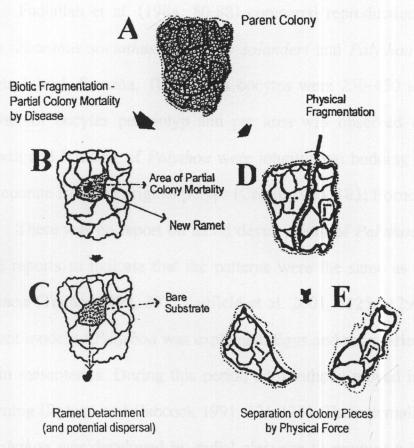


Figure 2 *Palythoa* schematic illustration of asexual reproduction by Fragmentation

Note: Fragmentation processes proceed from partial colony (top figure) tick arrow represent principle partway (A) partial colony, (B) biotic fragmentation (C) detachment o the ramet (D) physical fragmentation by substrate instability. (j' and j" are separate from partial colony as two boulders acting as substratum separate; storm activity might initiate this (C) ramet dispersal usually effected by current, slop of the bottom etc.

Source: Acosta, A., P. W. Sammarco, and L. F. Duarte. 2001. Asexual Reproduction in a zoanthid by fragmentation: the role of exogenous factors. **Bull. Mar. Sci.** 68: 369.

Fadlallah et al. (1984, 80-88) compared reproduction of three zoanthids, Zoanthus sociathus, Zoanthus solanderi and Palythoa caribaecum, in Galeta Island, Pamana. Their mean oocytes were 250-450 µm. The highest number of oocytes per polyp and per area was observed in Z. sociathus. Growth and budding of Palythoa were longitudinal budding from stolon but not separate from the original polyps (Cooke 1976, 183; Borneman 1998, 3).

There was no report on larval development of *Palythoa* but there were some reports to indicate that the patterns were the same as those of *Protopalythoa* (Tanner 1999, 397; Hadfield et al. 2001, 1125-1129). Larval development model of *Palythoa* was explained. Eggs and spermaries were developed in mesenteries. During this period zooxanthella moved into eggs before spawning (Ryland and Babcock 1991, 121-123). The externally fertilized egg of *Palythoa* was developed by radial cleavage to produce a hollow blastula. Gastrula formation took place by a mechanism to invaginate and proceed by a flattening of the blastula followed by progressive concave/convex deformation. After that the mouth and mesenteries began to form in the larva prior (Barnes et al. 1988, 68). Then after two weeks the larvae developed to the stage of zoanthella, and were able to settle after 17 to 18 days (Babcock and Ryland 1990, 239-236). The tentacles were formed later (Barnes et al. 1988, 68).

Growth rate

Animals usually grow in relation to their ages. But numbers in order Zoanthidea are different due to food risky, environmental disturbances and competition with other reef sessile organisms (Sebens 1977, 397-408; Karlson 1988a., 31) and strong wave (Koehl 1977, 437). Tanner (1997, 793-810; 1999, 390-397; 2000, 514-519) studied effect of density on the zoanthid by using Palythoa caesia in the Great Barrier Reef as a model. The model showed major differences in fission and fusion rates with density, with fission decreasing and fusion increasing as density increased. The growth rate of large colonies decreased substantially as density increased, although small colonies were unaffected. Mortality, on the other hand, was independent of density. Recruitment rates peaked at intermediate densities, but low overall. While density accounted for less than 10% of the variation in the demographic rates measured, it explained 62% of the variation in changes in cover within a given aggregation. Change in cover was greasted (30% in two years) for quadrats with 23-33% cover, and became negative for quadrats with over 80% cover. The highest density quadrats (90% cover) experienced an 18% decrease in cover over two years. When the data on colony level dynamics for each density were integrated into population dynamic models, he found a steady decrease in population growth rates with density, a long with an increase in the proportion of individuals in the largest size class. However, the proportional contribution of each size class to future generations (reproduction value) did not change whit density. These results suggest that the equilibrium

cover in the model was 84% and was reached in ≈ 40 years. To determine which density-dependent transitions were involved in population regulation, the strength of density dependence was varied in each independently. This sensitively analysis showed that only changes in probabilities of large colonies remaining large and producing medium colonies, were regulating. These results suggest that regulation is primarily acting on fission of large colonies to produce intermediate-size colonies, in combination with size specific growth rate. Fission rates decrease greatly with density, resulting in a greater proportion of large colonies at high densities and large colonies grow more slowly than small. Overall, this behavior was very similar to that of colonal plants which have a phalanx type life history. Although colony density change to produce intermediate-size depend on colonies size not species or population density.

Competition

Besides reproduction and growth ability, competition is the important point to make *Palythoa* widespread in many localities. Suckhnek and Green (1981, 619-684) found that *Palythoa* can overgrowth many reef sessile organisms such as *Acropora* spp., *Porites* spp., *Favia fragum*, *Millepora* spp., *Zoanthus solanderi*, *Haliclona* sp. and other unidentified sponges. Overgrowth in *Palythoa* can be divided in to 5 types:

1. Lateral aggression without physical contact, there exists a margin of dead tissues in front of the advancing Palythoa colony and it is assumed

that some from of allelochmical interaction is likely occurring (it possible that the *Palythoa* colony had previously advanced, smothering the coral polyps, then retracted, leaving a margin of bare space before advancing again) (Figure 3A).

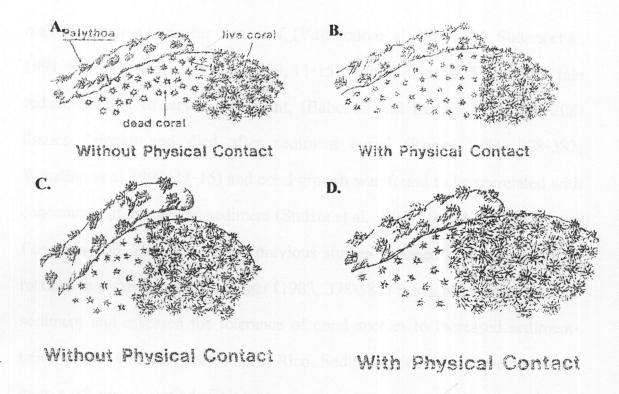
- 2. Lateral aggression with physical contact, the interaction may involve some tentacular action on the part of Palythoa or an actual smothering of subordinate species (Figure 3B).
- 3. Overtopping without physical contact, subordinates with zooxanthaellae would be shaded out of existence: it is likely that heterotrophic feeding would be severely impeded in is case as well No allelechemical or tentacular effect need be invoked. hear (Figure 3C).
- 4. Overtopping with physical contact, smothering would again prevail, eventually killing the subordinate. (Rapid growth rate would be necessary to over take the other species (Figure 3D).
- 5. Point settlement, settlement Palythoa directly onto a living organism (Figure 3E).

Karlson (1980, 894-898) reported that Palythoa was aggressive cnidarian overgrowing Zoanthus, P. porites, Agaricia, Siderastrea and Acropora as well as Millepora at an average rate of 1.3 cm/year (range = 0.3-3.5 cm/year). Bastidas and Bone (1996, 543-555) studied competition between two zoanthids Palythoa caribaeoum and Zoanthus sociatus in Isla Raton Cuerto Cabello, Venezuella, and found that P. carivaeom was strong competition compare to Z. sociatus. P. carivaeom did a greater the initial cover and growth rates were highly variable, ranging from 0.08 ± 0.02 to 0.12 ± 0.06 cm²

border/month and Z. sociatus raging from 0.05 + 0.15 to 0.14 + 0.51 cm² border/month. Hill (1998, 513-521) reported the spatial arrangement and genetic composition of populations of the zoanthid Parazoanthus parasiticus on sponge Callyspongia vaginalis. The pattern of dispersion of P. parasiticus on the C. vagimalis was randomly for four of six populations. Two populations exhibited overdispersion. Polyps density decreased towards the growing edge of C. vaginalis, and approximately 12% of polyps were in process budding. Electrophoretic variability at five enzyme loci indicated that population of P. parasiticus, cell collected from there species sponges, can consisted of genetically identical individuals. Heterozygosis at several loci indicated an absent of meiotic segregation, and therefore colonal reproducetion. P. parasiticus population may be initiated. One is that a single sexually derived planktonic larva settles on a sponge, and through asexually reproducetion colonizes all of the available surface area. Another possible is that a single genotype out completes a number of colonizers, and takes over the sponge surface through asexual reproduction. Finally, multiple colonization of same genotype may occur with subsequent asexual reproduction.

Environmental effect on Anthozoan behavior

Reef sessile organisms incur at various scales resulting from biotic and abiotic causes. (Wessling et al. 1999, 11-15) Sedimentation is cited as one of the



E. Point Settlement

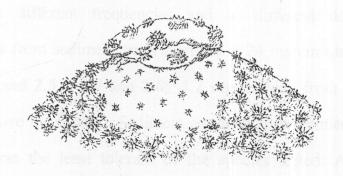


Figure 3 Type of overgrowth interactions *Palythoa* by growing over subordinate species

Source: Suckhnek, T. H., and D. J. Green. 1981. Interspecific competition between *Palythoa caribaeorum* and other sessile invertebrates on St. Croix Reefs, U.S. Virgin Island. In **Prog. 4**th. Int. Coral Reef Symp. Vol. 2. Manila:. University of Philippines, 682.

main destructive focus for coral reef, (Yamazato et al. 1987, 289; Sudara et al. 1991, 107-112; Wessling et al. 1999, 11-15) such as higher sedimentation rate reduce number of larvae settlement, (Babcock and Davies 1991, 205-208) tissues damage and died after sediment burial (Roger 1983, 378-382; Wessling et al 1999, 11-15) and coral growth was found to be correlated with concentrate of suspention sediment (Sudara et al. 1991, 107-112; Anthony and Febricius 200, 221-253). Some previous studies reported coral behaviour in relation to sediment effects. Roger (1983, 378-382) measured natural rates of sediment and assessed the tolerance of coral species to increased sedimenttation at San Critobal Reef, Puerto Rico. Sedimentation rates were measured over a 18 month period. Calcareous sediments were applied to colonies of Montastraea annularis, Diploria strigosa, D. clivosa, Acropora palmata and A. cervicornis in different frequencies and in different doses. Mean sedimentation rates from sediment traps were 96 ± 24 mg/cm²/day at 10 cm above the bottom and 2.5 ± 0.9 and 2.6 ± 1.2 mg/cm²/day from two sets of traps at 50 cm. above the bottom. Sediment application experiment indicated that A. palmate was the least tolerant of the species tested. Although A. cervicornis and D. stigosa colonies were not significantly affected, single applications of 800 mg/cm² to M. annularis colonies and of 200 mg/cm² to A. palmata colonies caused death of underlying coral tissue. Algae colonized portions of these corals. Sudara et al. (1991, 107-112) did comparison of growth of Porites lutea colonies at various sites around Khang Khao Island, on the Inner Gulf of Thiland. Growth studies of P.lutea revealed that horizontal growth was slower than vertical growth. Vertical growth of P. lutea

in the northern part of the island was faster during the period December to June when sedimentation was low levels of suspended solids were also higher during the period December to June. In the southern part of the island, coral growth was not as good during December to June as it was during July to December. Babcock and Davies (1991, 205-208) reported the effects of varying rates of sedimentation (0.5 to 325 mg/cm²/day) on settlement rates of Acropora millepora larvae were examined experimentally, in aquaria. Settlement and juvenile orientation onto conditioned cut coral plates were recorded after two days. Higher sedimentation rates reduced the number of settled larvae were not significantly affected by sedimentary regime. While total settlement was unaffected by under the conditioned experimental conditions, it likely to be reduced under field conditions, since accumulation of sediment on upward facing surfaces will greatly reduce the overall amount of suitable substratum available.

In *Palythoa*, Haywick and Mueller (1997, 36-46) reported that they incorporate sediment into their tissues as they growth. The amount of sediment assimilated is significant, averaging almost 45% of wet tissue weight. *Palythoa* assimilate all available minerals on the reef. There also no preference in terms of skeletal composition: coral grit, coralline red algae, *Halimeda* and other allochems are all equally assimilated into *Palythoa* tissues. The only preference is generally ≤ 125 μm in size, far finer than ambient sediment found adjacent to *Palythoa* colonies (predominantly > 500 μm) Acosta (2001, 119-120) suggested other sediment defense mechanisms of *Palythoa* on São Paulo coast, Brazil. During the winter, after period of

storms and continuous stress due to turbidity and an extremely high sedimentation rate, 19.7% of the total colonies monitored (n = 578) exhibited a thin layer of mucus covering all or part of the colony. Colonies remained retracted, with polyps close and feeding. Some of colonies stayed in this condition for weeks or even months into the winter, appearing dead. A reduction in height of the contracted polyps in the whole colony was evident during this stage, suggesting tissue resorption. Weeks later, the colony released this mucus layer, which was sometimes covered by algae and sediment grains, then opened their polyps and started to feed again. A shiny color on the surface characterized the colonies that finished this period of what might be termed diapause or dormancy. No colonies that experienced this stage died.

In addition to *Palythoa* mucus was composted by marine toxin call Palytoxin or PTX (Kimura et al. 1972, 611-617; Tosteson et al. 1995, 799-807; Gleibs et al. 1995, 1531-1537; Carte 1996, 271; Leech and Habener 1998, 1066-1072; Gleibs and Mebs 1999, 1521-1527; Acosta 2001, 125; Wu et al. 2003, 55). Plytoxin is considered to be one of the most compound, exhibition extreme in mammals (Tosteson et al. 1995,799-806; Gleibs et al. 1995, 1531-1537; Gleibs and Mebs 1999, 1521-1527) PTX was increased the cationic permeability of cell membranes and inhibits the (Na⁺, K⁺)-activated ATPase, effects that are completely reversed by ouabain. (Mebs 1998, 1519-1522; Rhodes et al. 2002, 631-636; Wu et al. 2003, 55-62) Palytoxin effect uses as an anti-perdition defense of *Palythoa*. (Hill 1998, 513)

CHAPTER 3

MATERIAL AND METHODS

Historical background of the Materials

Taxa and Background of Palythoa caesia

Kingdom

: Animal

Class

: Anthozoa

Subclass

: Zoantharia

Order

: Zoanthidea

Family

: Zoanthidae

Genus

: Palythoa

Species

: caesia

Diagnostic feature of Palythoa caesia (Burnett et al. 1997, 57-65)

Habitat

: Subtidal on coral reef. Intertridal on reef crests.

Generic featured

: sediment encrusted polyps buried in thick conenchyme.

From feature

: Polyps large (~ 8 mm diameter), colonies forming small

ovoid blobs approx. 10 cm. diameter or less. Generally

buff colored, ranging from almost white to dark brown.

Key to Great Barrier Reef and Torres Strait

l.	A Polyps sand encrusted2
	B Polyps not sand encrusted6
2.	A Polyps solitary. Found living on sublittoral sandy substrata, not coral
	reefs. Polyps very robust, rounded or slightly pointed at the base. Small
	specimens may be attached to small stones or sea grass blades by thin
	stalkSphenopus marsupialis
	B Polyps colonial, on coral reefs and rocky shores3
3.	A Polyps completely immersed in a thick coenenchyme when closeed.
	Colonies may be small (~3-10 cm) and ovoid coenenchyme with large
	polyps, or very large with small, densely packed polyps
	Palythoa caesia
	B Polyps free standing, conected only at base4
4.	A Intertidal. Disk green or brown, often with contrasting light colored
	mouth and hypostome. Polyps cylindrical when closed, with scapus
	and capitulum of equal diameter. Scapus may appear transversely
	wrinkled Protopalythoa mutuki
	B At extreme low water and subtidal5
5.	A Polyps fully open in shade. Disk uniform brown, mottled whit and
	brown/green, or brown with white radial lines. Often semi-closed in
	direct sunlight, with triangular appearance. Polyps trumpet shaped
	when closed Protopalythoa sp. 2

	Polyps white or sandy colored, partially buried in coral rock
6.	A Polyps large (to 70 mm), joined at base. Colonies usually with only
	a few polyps. Body wall with large tubercules usually arranged in
	longitudinal series. Intertidal and sublittoral Isaurus tuberculatus
	B Polyps small (rarely over 20 mm), without tubercles or immersed in
	a thick coenenchyme7
7.	A Polyps joined at base by stolones. Intertidal or sublittoral. Color
	highly variable, bright green, red, orange, yellow or brown. Oral disk
	uniform or platented, tentacles often strongly contrasting color disk
	Zoanthus coppingeri
	B. Polyps immersed in thick coenenchyme. Intertidal on exposed reef
	crests. Colonies forming extensive rubbery mats. Colonies strikingly
	coloured, powder blue or green Zoanthus vietnamensis

Study sites

P. caesia specimens and sedimentation rates were investigated at Khang Khao Island, inner part of The Gulf of Thailand, 13° 06′ 45″ N, 100° 48′ 52″ E. The Island is approximately 60 and 40 kilometers from Chow Phraya and Bangpogong river-mounts, respectively, so the area was effected by the sediment runoff during rainy season. (Ruengsawang 1999, 9) In the present study three stations of Khang Khao Island were selected for field study. All stations are influenced by the difference of monsoons, station A is

on the north coast of the island which is directly effected by Northeast monsoon during October to February. Station C is on a east coast of the island which is effected by Southwest monsoon during May to September and Southeast monsoon or Southeast trade during February to April. Station D is on the west coast of the island which is also effected by Southwest monsoon. The station descriptions are as follows: (Figure 4)

Station A divided into 2 zones as;

As The shallow zone

Ad The deep zone

Station C divided into 2 zones as:

Cs The shallow zone

Cd The deep zone

Station D divided into 2 zones as;

Ds The shallow zone

Dd The deep zone

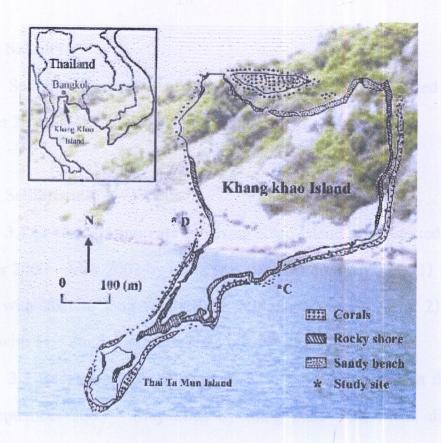


Figure 4 Map of the study area

Field Experiments

Measurement of the environmental factors

1. Temperature

Surface seawater temperature (SST) of Khang Khao Island was detected during September 2000 – May 2002 by using a digital underwater thermometer.

2. Salinity

Salinity of seawater of Khang Khao Island was detected during September 2000 – May 2002 by using a salinometer.

3. Sedimentation

- 3.1 Sedimentation rate of Khang Khao Island was detected during September 2000 May 2002 by using two sizes of sediment traps, 1) 1.5 cm. diameter with 20 cm. long (Suthanaruk 1983) (Figure 5B.) and 2) 5 cm. diameter with 11.5 cm. long (English et al. 1997, 59) (Figure 5A.).
- **3.2 Resuspension sediment** of Khang Khao Island was detected during September 2000 May 2002 by using plate with 9 cm. diameter (Figure 5C.).

4. P. caesia specimen

From September 2000 – May 2002, three colonies per station were marked and measured continuously by using underwater photography to detect growth and mortality rates. Nine *P. caesia* colonies from 3 stations (Cs, Ds and Dd) were randomly collected by a sharp knife and fixed in 10% formalin: seawater to detect gamete development and sediment in the tissues. For *P. caesia* interactions, three belts transects and 1 m² quadrates, (Aerts and Soest 1997, 126-128) were applied randomly along the reef bed. Four categories of interactions were defined.

- 1) Aggregation without physical contact
- 2) Aggregation with physical contact

- 3) Overtopping
- 4) Point settlement (Suckhnek and Green 1981, 679-684)



Figure 5 Study of sedimentation and resuspension rate by three types of sediment traps

Note: A. sedimentation trap 5 cm. diameter B. sedimentation trap 1.5 cm. diameter and C. sediment plate.

Laboratory works

1. Sediment trap

To estimate the sediment dry weigh, dry and clean beakers were pre-weighed and running numbers (Figure 6).



Figure 6 Washing sediment traps by running water



Figure 7 Place beaker in room temperature



Figure 8 Dry sediment from a sediment trap at 105°C, 24 hour

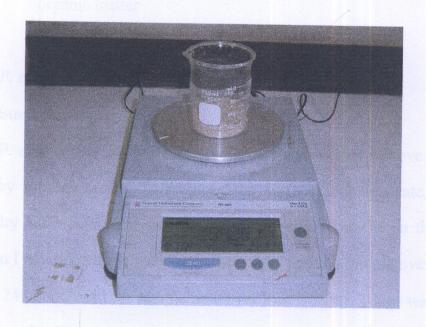


Figure 9 Weighing



Figure 10 Dry sample burn in muffle furnace at 550°C, 5 hours to remove organic matter

2. P. caesia colonies

Surface sediment

P. caesia specimens were note surface area before remove surface sediment by running water. To estimate the surface sediment rate, I preweight a dry filter paper and poured the water with sediment on the filter paper. Then I determined the weight of filter paper and the sample oven dry at 105°C for 24 hours (until the sample no longer lose weight). Tongs were used for removing the filter paper with sample from the oven and placed it on a desiccator containing silicagel and allowed the sample and filter paper to cool at room temperature and weighed (Figure 11).

Surface sediment of *P. caesia* colony was

Filter paper with sediment weigh - Filter paper weigh = Sediment weigh



Figure 11 Sediment from P. caesia surface on a filter paper

3. Entomb sediment

A piece of 2 x 3 cm. from each *P. caesia* sample was cut with a sharp dissection knife then dried at 105°C, 24 hours or until the sample no longer lose weigh before weighed (DW). Sample placed in crucibles and weighed (CW) before ash in muffle furnace at 550°C, 6 hours to investigate lose organic weigh. Tongs were used for removing the crucible from muffle furnace and place in a dedicator contain silica gels, and allowed the sample and crucible cool at room temperature and weighed (IW).

Sediment from sediment traps were remove to beaker respectively by running water and made the sediment traps labeled on this. Place beaker in room temperature for sediment were settling for 3-4 days. (Figure 7) Poor slowly top water (no sediment) dry at 105°C, 24 hour or until the sample no longer lose weight. (Figure 8) Tongs were used for removing the beaker from the oven and placed it on a desiccator containing silicagel and allowed the sample and beaker to cool at room temperature and weighed.

Then 10 mg from each samples were pick to running crucibles (crucibles were pre-weight before add the sample) respectively. After that I place the crucible in a muffle furnace at 550°C for 5 hours to destroy the organic matter. (if black charcoal deposit are still visible, then continues ashing) The temperature did not exceed 550°C because the CaCO₃ would be converted to CaO₂, thus resulting in a biased ash sample free dry weight. Finally the furnace has been allowed to cool for several hours. I removed the curable containing the ashed samples and cooled them at room temperature in a desiccator and then weight (Wa).

Organic mater of sediment

10 - Wa = Organic matter

Organic and inorganic weigh of P. caesia.

CW - DW = Inorganic weigh (IW)

DW - IW = Organic weigh

Histological Study

In order to examine gamete development, histological sections of marked colonies were prepared 1-2 polyps of *P. caesia* were separated by a sharp dissection knife. After desilified in 20% hydrofluoric acid for 24 hours and decalcified in a solution compared equals parts of formalin and saturation formic acid made up 1:9 with water (Ryland and Babcock, 1991, 118). The tissues were washed in running water to remove formalin then dehydrated through a graded series of ethanol: 70%, 80%, 90%, 95% and absolute 1, absolute 2 for 30 minutes, respectively. The samples were transferred to ethanol – xylene (1:1), xylene 1, xylene 2 for 30 minutes respectively, then passed to be impregnated in warm xylene – paraffin (1:1), pure paraffin 1, pure paraffin 2 and finally embedded in pure paraffin in plastic block. The samples were cut into serial section, (10µm thick) and stained with hematoxylin and eosin. Both transverse and longitudinal sections were then examined with a light microscope for reproductive structures.

CHAPTER 4 RESULTS

Environmental Factors

Salinity

The present study recorded salinity at Sichang Island, Chonburi Province from February 2000 – December 2001 as follows:

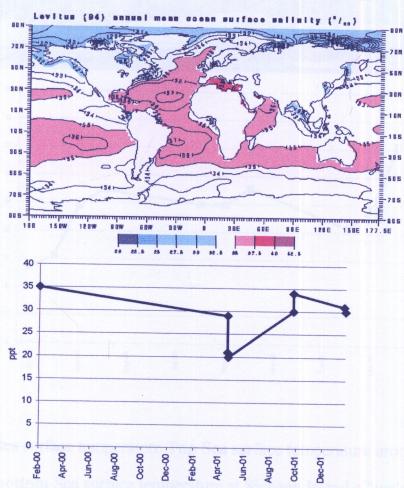


Figure 12 Salinity **Top** Salinity around The World **bottom** salinity during February 2000 – December 2001 at Sichang, Chonburi Province

Sea Surface Temperature

The data of sea surface temperature at Sichang Island, Chonburi Province during January 2000- May 2001 are showed as follows:

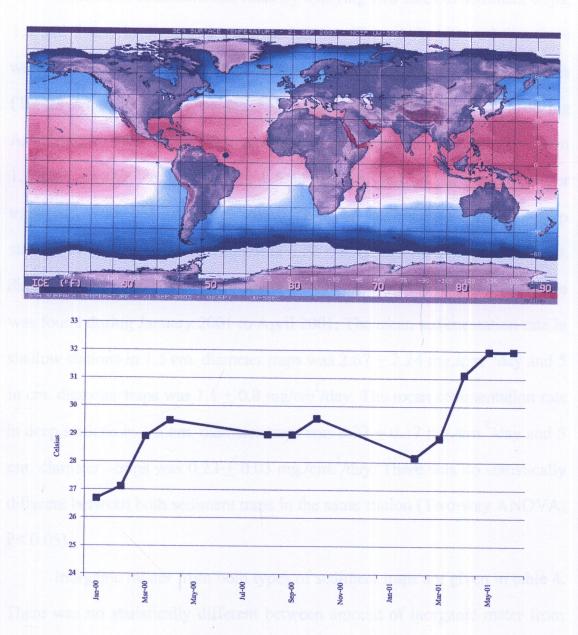


Figure 13 Sea surface temperature **Top** Sea surface temperature around the word **bottom** Sea surface temperature at Sichang Island, Chonburi Province during

January 2000- May 2001

Source: Geo-Information and Space Technology Development Agency

Sedimentation rates and Resuspension of bottom sediment

Studies of sedimentation rates by applying two sizes of sediment traps, 1.5 and 5 cm. in diameters were carried out. The results showed that there were higher mean sedimentation rates in shallow stations than deep stations (Tables 2 and 3 Figures 14 and 15). Sedimentation rates were high during April 2001 to July 2001. The mean sedimentation rate in shallow stations in 1.5 cm. diameter – traps was 7.11 ± 5.64 mg./cm. ²/day and in 5 cm. diameter traps was 5.07 ± 1.71 mg/cm²/day. The mean sedimentation rate in deep stations in 1.5 cm. diameter traps was 5.92 ± 8.41 mg./cm. ²/day and in 5 cm. diameter traps was 2.67 ± 2.24 mg./cm. ²/day. The lowest sedimentation rate was found during January 2001 to April 2001. The mean sedimentation rate in shallow stations in 1.5 cm. diameter traps was 2.67 ± 2.24 mg./cm. ²/day and 5 in cm. diameter-traps was 1.1 ± 0.8 mg/cm²/day. The mean sedimentation rate in deep stations in 1.5 cm. diameter traps was 0.32 ± 0.17 mg./cm.²/day and 5 cm. diameter -traps was 0.23 ± 0.03 mg./cm. day. There was no statistically different between both sediment traps in the same station (Two-way ANOVA, P < 0.05)

Inorganic matter from both types of sediment traps are given in table 4. There was no statistically different between amount of inorganic mater from both traps. Sedimentation rates and inorganic matter had a relationship with mortality rate of *P. caesia*. If sedimentation rate was increased, mortality rate of *P. caesia* was increased as well.

Resuspension of bottom sediment was studied by using sediment plates during April 2001 to May 2002. The results are showed in Table 6 and Figure 16. The results revealed that there were high resuspension of bottom sediment rates in shallow stations than deep stations. The highest resuspension of bottom sediment rate was found in October 2001. The mean resuspension of bottom sediment rate was $545 \cdot 1.91 \pm 1.33 \cdot \text{mg./cm.}^2/\text{day}$ and the lowest resuspension of bottom sediment rate found in September 2001. The mean resuspension of bottom sediment rate was $0.52 \pm 0.52 \cdot \text{mg./cm.}^2/\text{day}$. There was statically different between sedimentation rates by sediment traps and resuspension of bottom sediment by sediment plates in the same station (Twoway ANOVA, P<0.05).

Table 2 Sedimentation rates in sediment traps 1.5 cm. diameter (mg./cm.²/day.)

			SF	SHALLOW STATIONS	STATIO	NS			·	DEEP ST	DEEP STATIONS		
		Jul/00-	Jul/00- Sep/00- Jan/01- Apr/01- Jul/01- Oct/01-	Jan/01-	Apr/01-	Jul/01-	Oct/01-	Jul/00-	Sep/00-	Jan/01-	Jul/00- Sep/00- Jan/01- Apr/01- Jul/01- Oct/01	Jul/01-	Oct/01-
		Sep/00	Sep/00 Jan/01 Apr/01 Jul/01	Apr/01	Jul/01	Oct/01	May02	Sep/00	Sep/00 Jan/01 Apr/01	Apr/01	Jul/01	Oct/01 Mav02	May02
A	A mean	1.454	0.937	0.367	5.268	2.162	1.086	3.590	0.449	0.225		0.703	0.405
	SD	0.137	0.172	0.107	2.849	0.716	0.424	3.590	0.185	0.063	12.221	0.291	0.100
C	mean	2.795	0.269	0.386	8.518	8.518 11.545	6.288	1.544	0.287	0.000	1.544	1 893	CTT C
	SD	0.125	0.009	0.113	5.434	2.418	2.999	0.313	0.067	0.000	0.313	0.853	1 871
Ω	mean	4.512	0.292	0.237	966.9	2.162	0.000	2.269	0.333	0.470	4.248	0.931	1.971
	SD	1.957	0.030	0.033	7.662	0.696	0.000	0.561	0.154	0.197	3,111	0.322	203.0
				***************************************							•	111	2000

Table 3 Sedimentation rates in sediment traps 5 cm. diameter (mg./cm.²/day.)

			SH	IALLOW	SHALLOW STATIONS	NS				DEEP STATIONS	ATIONS		
		Jul/00-	Jul/00- Sep/00- Jan/01- A	Jan/01-	Apr/01-	Jul/01-	.pr/01- Jul/01- Oct/01- Jul/00- Sep/00- Jan/01- Apr/01- Jul/01-	Jul/00-	Sep/00-	Jan/01-	Apr/01-	Jul/01-	Oct/01-
		Sep/00	Sep/00 Jan/01 Apr/01	Apr/01	Jul/01	Oct/01	Jul/01 Oct/01 May02 Sep/00 Jan/01 Apr/01 Jul/01 Oct/01	Sep/00	Jan/01	Apr/01	Jul/01	Oct/01	May02
A	A Mean	6.728	1.349	0.100	1.063	0.287	0.000	0.239	0.539	0.127	1.158	1.158 0.428	0.875
	SD	1.087	0.094	0.000	0.451	0.237	0.000	0.058	0.165	0.033	0.496	0.263	0.815
C	Mean	5.574	0.270	1.591	2.587	0.251	2.769	0.000	0.000	0.000	1.686	1.476	1.404
	SD	1.150	0.042	0.133	1.449	0.133	1.041	0.000	0.000	0.000	2.009	1.457	0.978
Q	D Mean	4.487	0.130	0.160	1.975	0.672	1.804	5.337	0.660	1.230	4.047	0.999	1.717
	SD	1.336	0.000	0.000	2.172	0.300	1.765	0.985	0.000	0.000	2.121	0.234	1.783

Table 4 Mean amount of inorganic mater from sediment traps (g./ash)

			SE	IALLOW	SHALLOW STATIONS	SN				DEEP ST	DEEP STATIONS		
		Jul/00-	Jul/00- Sep/00- Jan/01- Apr/01- Jul/01- Oct/01-	Jan/01-	Apr/01-	Jul/01-	Oct/01-	Ju1/00-	Sep/00-	Jan/01-	Sep/00- Jan/01- Amr/01- 111/01-	F11//01-	Oct/01-
		Sep/00	Sep/00 Jan/01 Apr/01	Apr/01	Jul/01	Oct/01	Oct/01 Mav02	Sen/00	Sen/00 Ian/01	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	1101		-10/20
4	Mean	Mean 0.050	0.960	9760	-			and	omin of		Jul/UI	Octol May02	May02
1	Thoras I	0	0.009	0.702	1.000	0.949	0.813	0.840	0.911	0.846	0.987	0.953	0.887
	S.D.	0.030	0.169	0.258	0.000	0.074	0.317	0.131	0.907	0.000	0.041	0.139	0.160
C	Mean	0.932	0.907	0.908	1.000	0.991	0.816	0.900	0.895	0.000	1,000	0.078	0.053
	S.D.	0.004	0.039	0.057	0.000	0.028	0.273	0.067	0.007	0.000	0000	0.045	0.03
Ω	Mean	0.948	0.717	0.850	1.000	1.000	0.945	0.948	0.907	0.888	1.000	0.953	0.957
	S.D.	0.020	0.481	0.026 0.000	0.000	0.000	0.012	0.018	0.081	0.093	0000	0.103	0.000
									())		200.0	0.10	0.012

Table 5 Mean amount of organic matter from sediment traps (g./DW.)

			SH	SHALLOW STA	STATIONS	NS				DEEP S	DEEP STATIONS	100	
		Jul/00-	Sep/00-	Jan/00-	Jul/00- Sep/00- Jan/00- Apr/01- Jul/01- Oct/01- Jul/00- Sep/00- Jan/00- Apr/01- Jul/01-	Jul/01-	Oct/01-	Jul/00-	Sep/00-	Jan/00-	Apr/01-	Jul/01-	Oct/01-
		Sep/00	Sep/00 Jan/00 Apr/01 Jul/0	Apr/01	Jul/01	Oct/01	Oct/01 May02	Sep/00	Sep/00 Jan/00 Apr/01	Apr/01	Jul/01	Oct/01	May02
A	A Mean	0.050	0.131	0.235	0.000	0.073	0.187	0.170	0.089	0.186	0.000	0.094	0.113
	S.D.	0.030	0.169	0.258	0.000	0.080	0.317	0.170	0.028	0.035	0.000	0.191	0.160
C	Mean	0.070	0.093	0.092	0.000	0.090	0.184	0.100	0.105	0.000	0.000	0.062	0.047
	S.D.	0.004	0.039	0.057	0.000	0.000	0.273	0.067	0.007	0.000	0.000	0.058	0.044
Ω	Mean	0.052	0.283	0.150	0.000	0.000	0.055	0.056	0.093	0.134	0.000	0.079	0.043
	S.D.	0.019	0.481	0.026	0.000	0.000	0.012	0.014	0.099	0.084	0.000	0.125	0.020

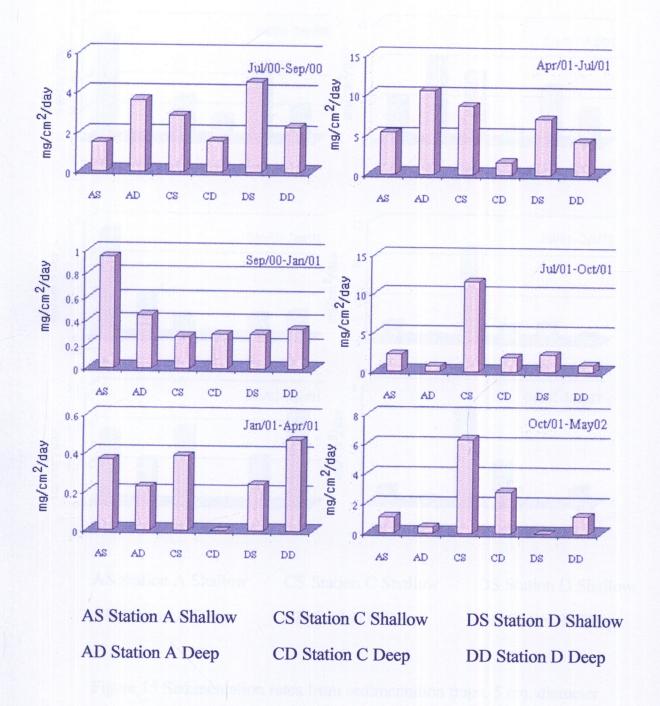


Figure 14 Sedimentation rates from sedimentation traps, 1.5 cm. diameter

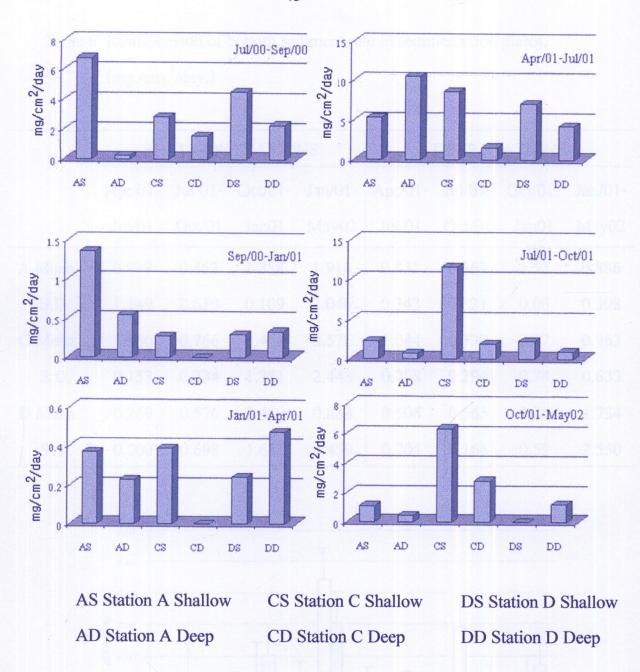


Figure 15 Sedimentation rates from sedimentation traps, 5 cm. diameter

Table 6 Resuspension of bottom sediment rate in sedimentation plates. (mg./cm.²/day.)

	CII	ALLOW	CT A TIO	NIC	,	DEED CT	CATIONIC	
	SH	ALLOW	STATIO			DEEP SI	TATIONS	
	Apr/01-	Jul/01-	Oct/01-	Jan/01-	Apr/01-	Jul/01-	Oct/01-	Jan/01-
	Jul/01	Oct/01	Jan01	May02	Jul/01	Oct/01	Jan01	May02
A Mean	0.929	0.462	1.558	1.916	0.431	0.861	1.53	0.886
S.D.	1.149	0.610	0.109	2.046	0.343	0.821	0.06	0.398
C Mean	1.160	0.766	3.448	2.576	1.044	0.330	0.87	0.963
S.D.	0.153	0.234	1.781	2.448	0.708	0.294	0.74	0.633
D Mean	0.269	0.576	2.489	0.883	0.506	0.565	1.26	1.734
S.D.	0.000	0.698	1.627	0.459	0.704	0.166	0.59	2.550

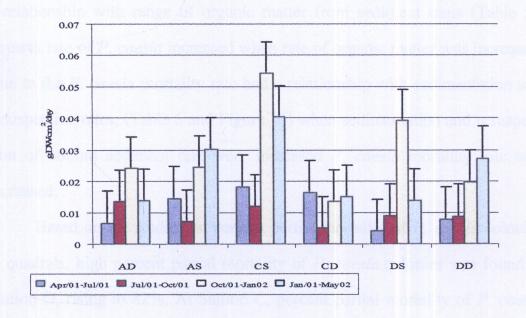


Figure 16 Resuspension of bottom sediment rates from sedimentation plates

Growth and Mortality rates, Change of colonies size and Percent partial Mortality of *P. caesia*

Growth and mortality rates of P. caesia from underwater photographs during January 2000 - June 2001 are given in Figure 18. The mean growth and mortality rates of P. caesia at Station D were higher than those at Station C. The mean growth rates were 1.81 ± 1.06 cm./mo. and 1.08 ± 1.49 cm./mo respectively. The mean mortality rates were 3.32 ± 2.74 cm./mo. and 1.61 ± 1.52 cm./mo, respectively. There was no statically different between growth and mortality rates of P. caesia at the same station and in a period of time. (ONEWAY-ANOVA P< 0.05) (Figure 17) Regarding to effects of environmental factors on growth and mortality rates of P. caesia, the growth rate had a relationship with range of organic matter from sediment traps (Table 5). Growth rate of P. caesia increased when rate of organic matter was increased. Due to the P. caesia mortality rate had a relationship with sedimentation and resuspension of bottom sediment rate were increased P. caesia mortality rate was increased.

Based on the studies on percent partial mortality of *P. caesia* colonies in quadrats, high percent partial mortality of *P. caesia* colonies was found at Station D, rising to 82%. At Station C, percent partial mortality of *P. caesia* colonies was only 25%. This result was agreeable with mortality rate of *P. caesia* studied by using underwater photographs.

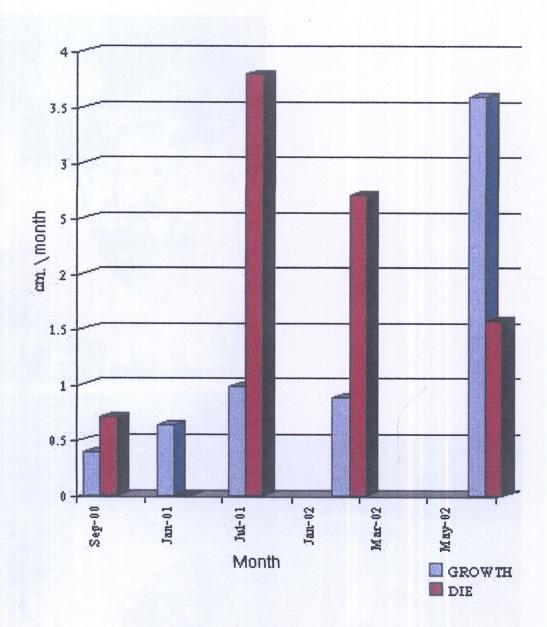


Figure 17 Growth and mortality rates of P. caesia

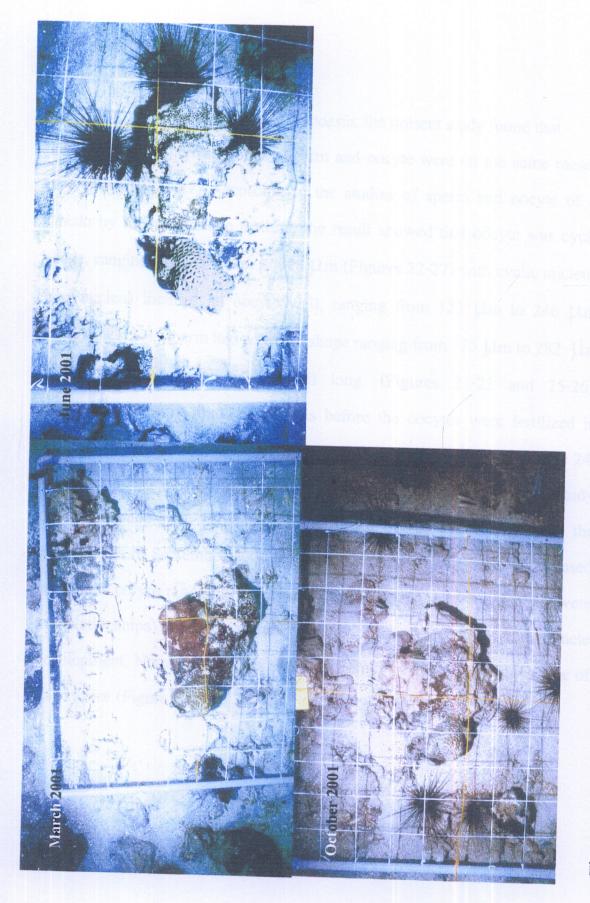


Figure 18 Growth rate, mortality rates and changes in colony sizes in the filed study

Sexual reproduction of P. caesia

For sexual reproduction of P. caesia, the present study found that P. caesia was a hermaphrodite. Sperm and oocyte were on the same mesentery. (Figures 21-27) According to the studies of sperm and oocyte of P. caesia by microtechnique method, the result showed that oocyte was cyclic shape, ranging from 246 μ m to 498 μ m (Figures 22-27) with cyclic nucleus. The nucleus inclined an oocyte wall, ranging from 123 μ m to 246 μ m. (Figures 21-27). Sperm had an ovoid shape ranging from 123 μ m to 282 μ m with 794 μ m to 2,706 μ m tail long. (Figures 21-23 and 25-26). Zooxanthellae were found in oocytes before the oocytes were fertilized in water column because the larvae in their tissues were not found. (Figure 24) P. caesia produced gonads throughout the year. Although during the study period. I found high percentage of fertilized polyps per colony from the observations in September 2000 - January 2001, 94.31% (Figure 19). Based on the 650 polyps randomly detected under compound microscope, there were no relationships of polyp size and polyp position in colony and gamete development. Number of polyps showed strongly relationship with change of colony size (Figure 20).

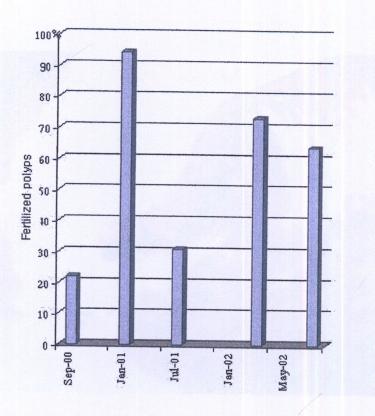


Figure 19 Percentage of fertilized polyps from each sampling periods

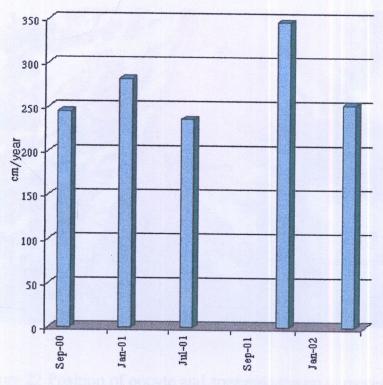


Figure 20 Change of colonies size in P. caesia

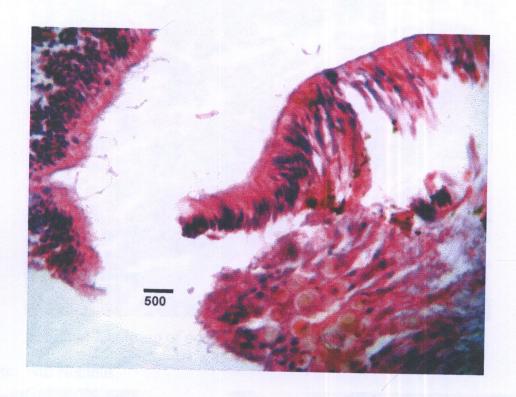


Figure 21 Oocyte and Spermatozoa on the same gonad

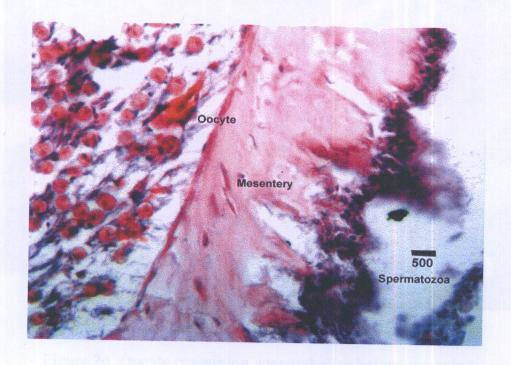


Figure 22 Position of oocyte and spermatozoa on a mesentery

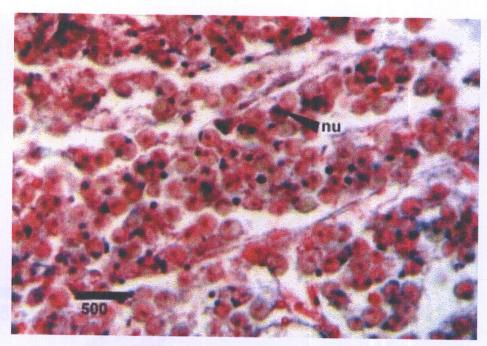


Figure 23 Group of oocytes in gonad and nucleus in oocytes

Note: Nu. Nucleus.

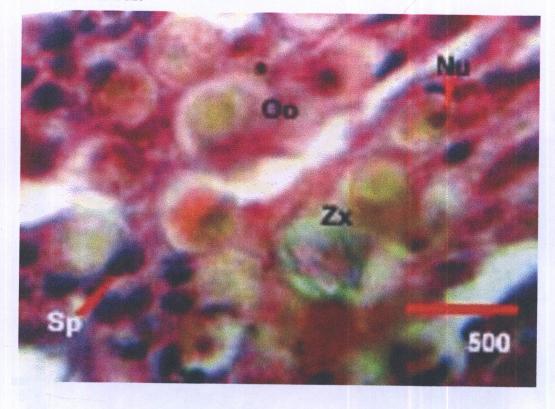


Figure 24 Oocyte containing zooxanthellae before spawning

Note : Sp = Spermatozoa, Oo. = Oocyte, nu = Nuclease, Zx = Zooxanthella

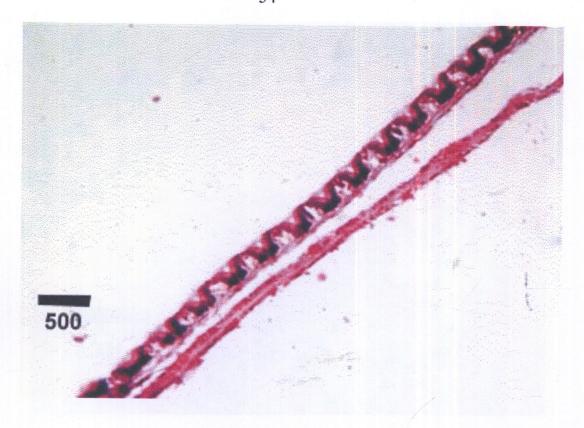


Figure 25 Sperm in a histological section

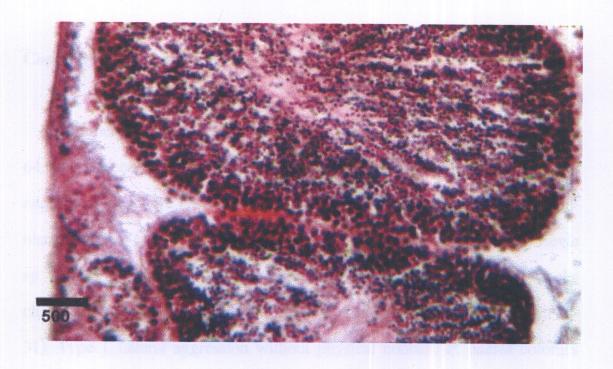


Figure 26 Mature spermatocyte found in a sample

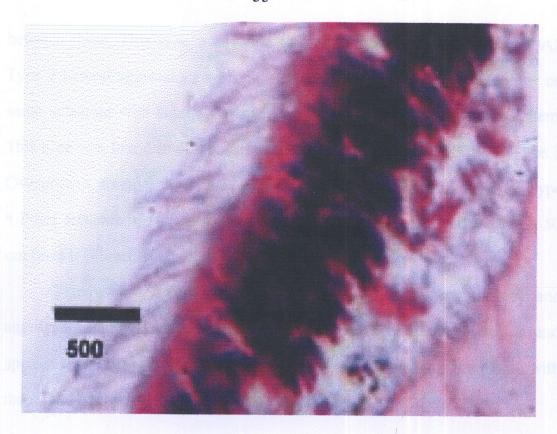


Figure 27 Spermatozoa and sperm tail in a histological section

Competition of P. caesia

Settlement of *P. caesia* was found on four types of substrates, rock 64.9%, dead coral 18.9%, sand with rubber 10.8% and living coral 5.4% *P. caesia* colonies were observed in competition with 6 species of sessile organisms, *Porites lutea* 46.4%, *Symphyllia* sp. 3.6%, *Zoanthid* sp. 3.6%, *Favia* sp. 3.6&, *Palythoa* sp. 3.6% and *Haliclona* cf. *delicatula* 39.2%. The results clearly showed that there were four types of aggregations (Table 7 Figures 28-31). Type 1 Lateral aggression without physical contact. *P. caesia* colonies grew on the same substrate with sessile organisms but no tissues in contact

between them. This type was found only with *Porites lutea*. (Figure 30A). Type 2 Lateral aggressions with physical contact, *P. caesia* colonies grew on same substrate with sessile organisms and tissues in contact between them. This type was found in all species at the study sites. (Figures 30B-C) Type 3 Overgrowth, found only with *Porites lutea* and *Haliclona* cf. *delicatula*. Type 4 Point settlement, *P. caesia* grew without neighbor reef sessiles around 50 cm from it (Figure 31C).

The result showed high level of *P. caesia* settlement on rock and aggregation with *Porites lutea* because they were dominant substrate and species in the study suites. In certain cases, *P. caesia* was in aggregation with more than one types per colony.

Distribution Patterns of P. caesia

Studies on distribution patterns of *P. caesia* by using at stations A, C and D revealed that the highest population density of *P. caesia* was at station D was percent cover 5.76%. The distribution pattern of *P. caesia* at station D was randomly distributed at depths 2.8 m to 3.6 m, 2 m to 20 m from the shoreline of Khang Khao Island (Figure 32). At station C, percent cover was 5.58%, and the distribution pattern of *P. caesia* was also randomly distributed at depths 2 m to 5 m, 13 m to 23 m from the shoreline of the island (Figure 33). *P. caesia* was also found at station A but the population density was relatively low.

Table 7 Percent of aggregation of P. caesia with other species.

	contact	without	overgrowth	%total
		contact	o voigio vuii	interaction
Porites lutea	62.23	15.38	15.38	46.42
Symphyllia sp.	100			3.57
Zoanthid sp.	100			3.57
Favia sp.	100			3.57
Palythoa sp.	100			3.57
Haliclona cf. delicatula	27.27		72.72	39.28
				100

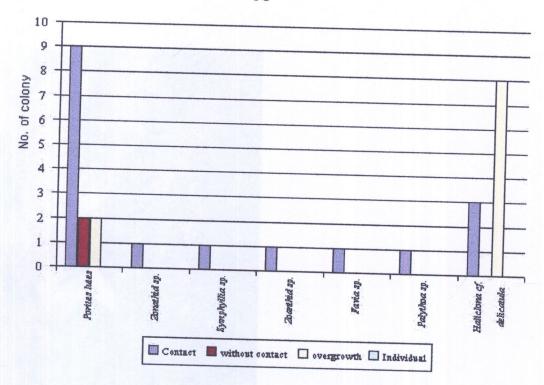


Figure 28 Number of sessile colonies in aggression with P. caesia

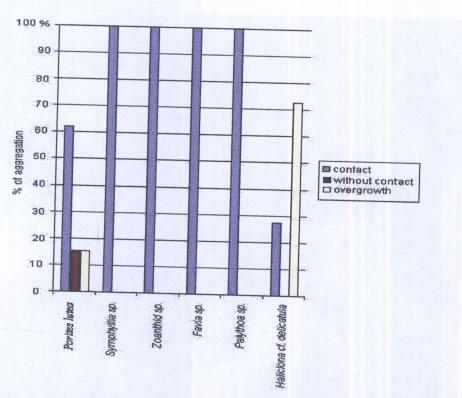


Figure 29 Percent of aggregation of P. caesia with other reef organisms

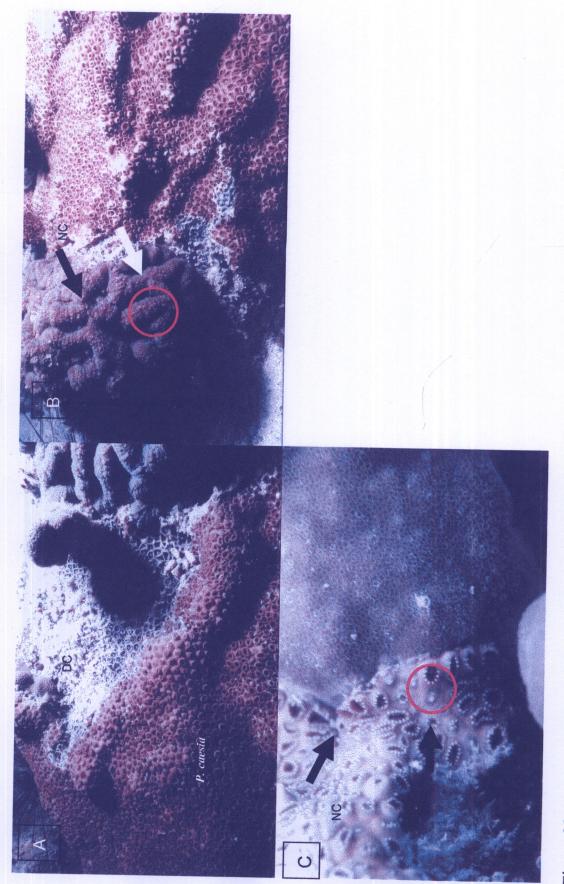


Figure 30 A. P. caesia tissues grew on dead coral without contact B.-C. (C) P. caesia damaged some areas of other species

Note: (NC) with living coral

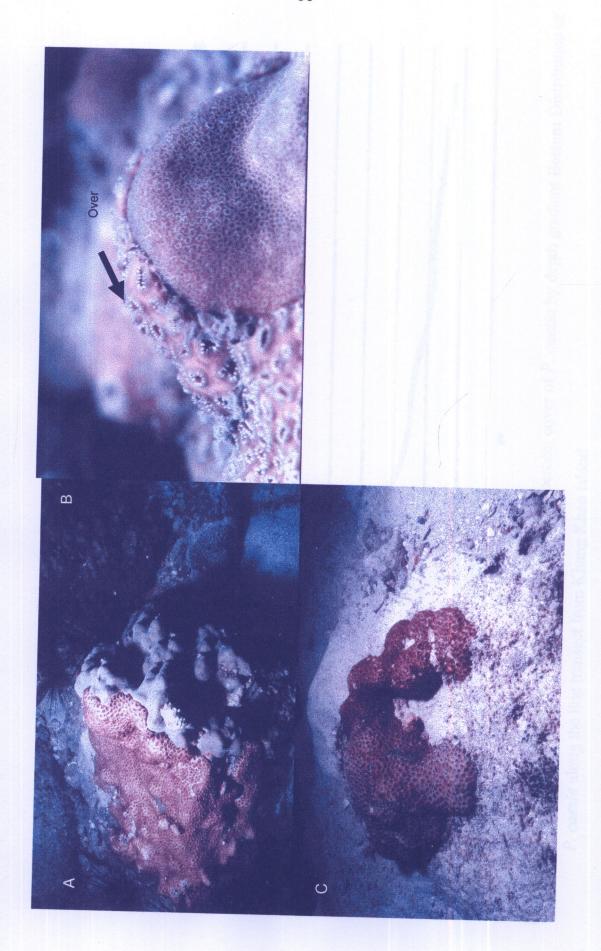


Figure 31 A. -B. P. caesia tissues overgrew on living coral. C. P. caesia colony settlement on rock without neighbor coral

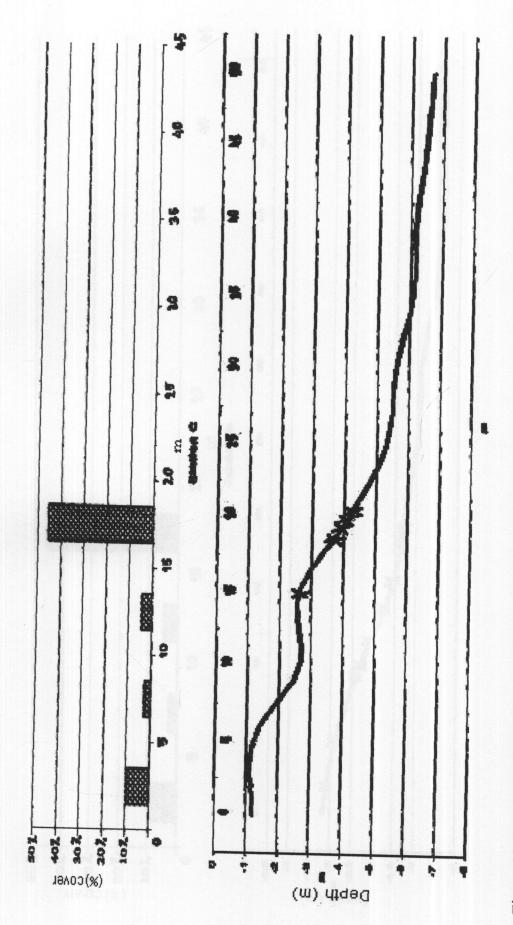


Figure 32 Distribution patterns of P. caesia on station C Top percent cover of P. caesia by depth gradient Bottom Distribution of P. caesia along the line transect from Khang Khao Island

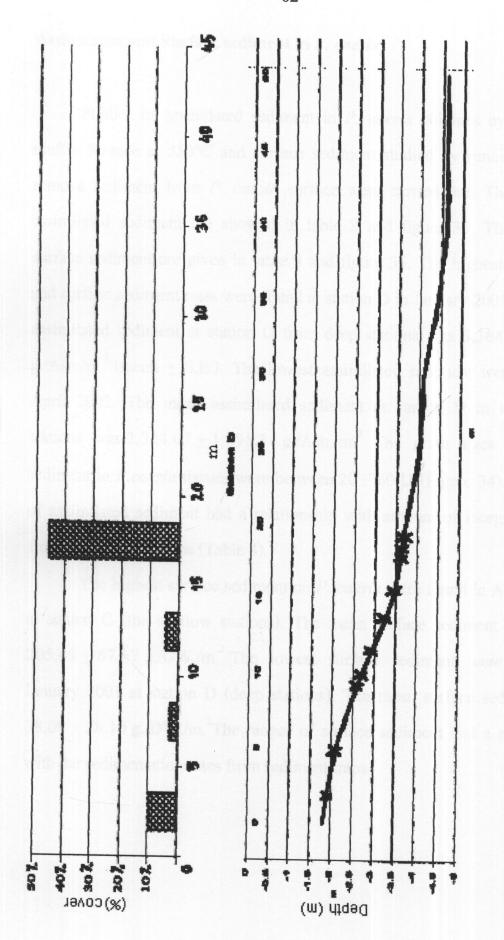


Figure 33 Distribution patterns of P. caesia on station D Top percent cover of P. caesia by depth gradient Bottom Distribution of P. caesia along the line transect from Khang Khao Island

Assimilated and Surface sediment in P. caesia.

Studies of assimilated sediment in *P. caesia* colonies by burning in muffle furnace at 550° C and surface sediment studied by running water to remove sediment from *P. caesia* surface were carried out. The results of assimilated sediment are showed in table 8 and figure 35. The results of surface sediment are given in table 9 and figure 36. The highest assimilated and surface sediment rates were found at station D in January 2001. The mean assimilated sediment at station D from deep stations was $3,364.8 \pm 806.86$ g./Ash./m.² (mean \pm S.E.). The lowest assimilated sediment were found in April 2002. The mean assimilated sediment at station D in the shallow stations was $2,324.07 \pm 1,095.34$ g./Ash./m.² The grain sizes of entomb sediment in *P.caesia* tissues were between 20 -760 μ (Figure 34). The range of assimilated sediment had a relationship with amount of inorganic matter from the sediment traps (Table 4).

The highest surface sediment on P. caesia were found in August 2000 at station C (the shallow stations). The mean surface sediment rates were 205.49 ± 87.67 g./DW./m.² The lowest surface sediment were found in January 2001 at station D (deep stations). The mean surface sediment was 38.09 ± 28.10 g./DW./m.²The ranges of surface sediment had a relationship with the sedimentation rates from sediment traps.

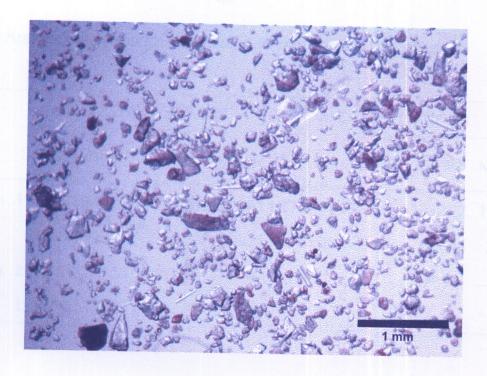
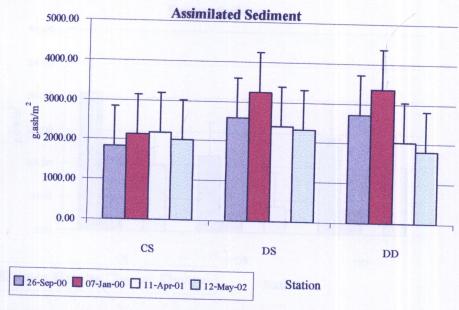


Figure 34 Entomb sediment found in P. caesia tissues

Table 8 Amount of assimilated sediment in P. caesia tissues (g./Ash./m.²)

	SH	ALLOW	STATIO	NS		DEEP ST	CATIONS	\$
	Sep-00	Jan-01	Apr-01	May-02	Sep-00	Jan-00	Apr-01	May-02
C Mean	1834.57	2146.30	2192.59	2016.67	2714.81	3364.81	2032.72	1804.17
S.D.			2322.25		12.3 81	4192.60	542.44	257.62
D Mean	2582.72	3247.53	2396.83	2324.07	3125	2 5 10	0.84	13.24
S.D.	1533.09	2666.26	1095.35	633.72	-	_	_	-



CS station C at shallow

DS station D at shallow

DD station D at deep

Figure 35 Amount of assimilated sediment in P. caesia tissues

Table 9 Amount of surface sediment on P. caesia colony (g./DW./m.²)

	SH	ALLOV	V STATIO	ONS		DEEP S	TATION	S
	Sep-	Jan-	Apr-	May-	Sep-	Jan-	Apr-	May-
To the second	00	00	01	02	00	00	01	02
C Mean	205.49	21.10	31.62	99.22	143.88	38.09	90.22	101.55
S.D.	87.67	7.87	22.15	75.18	34.25	28.10	0.84	13.24
D Mean	113.44	17.69	147.24	41.04	-	-	at/Khan	g Klaio -
S.D.	33.08	10.94	94.14	3.58	- 2	49 0 32/0	io, respe	ctively.

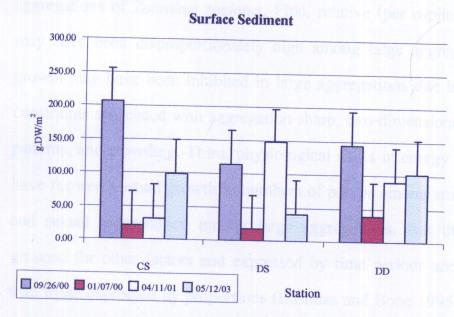


Figure 36 Amount of surface sediment on P. caesia colony

CHATER 5

DISCUSSION

Growth and Mortality rates

The results of growth and mortality rates of P. caesia at Khang Khao Island, Chonburi Province was 1.45 cm/month and 2.46 cm/mo, respectively. Karlson (1988b, 1225) explanations for high relative growth rates of small aggregations of Zoanthus sociatus. First, relative (per capita) mortality rates may have been disproportionately high among large aggregations. Second, growth may have been inhibited in large aggregations due to morphological constraints associated with aggregation sharp, two-dimensional clonal growth patterns, and crowding. Third, physiological shifts in energy allocations may have favored asexual growth in numbers of polyps among small aggregations and sexual reproduction among large aggregations. But the growth rates grouped for other factors and expressed by time periods are slightly higher than those expressed by proportions (Bastidas and Bone 1996, 550). There is evidence that growth form and rates can vary directly in response to limited food and space (Karlson 1988b, 1229). The results showed that growth rate of P. caesia had a relationship with range of organic matter from sediment traps. Anthony and Fabricius (2000, 221-253) have supported the idea of sediment as a food source. They showed that suspended sediment had a lower food value than zooplankton or phytoplankton. It constituted an often more

abundant component of the session in aquatic environment. It may be the predominant food source for suspension feeders, especially in turbid nearshore habitats.

Mortality rate of *P. caesia* has an effect from high sedimentation rate. This result can be answed by sedimentation rate of sediment traps. The result is agreeable with the study of Wesseling et al. (1999, 11-18) and Roger (1983, 378-382) as noted in the literature review. The other causes of mortality and partial mortality are from predator or diseases (Gleibset et al. 1995, 1531; Meb 1998, 1521; Gleibs and Mebs 1999, 1521; Acosta 2001, 131). The diseased colony is perceptible one or some polyps surrounding polyps appeared swollen bumpy morphology. The disease did not appear to produce excessive mucus as a sediment defense mechanism, when compared to healthy colony (Acosta. 2001, 117). The diseases can cause a partial mortality of *Palythoa* colony and resulting in separation to fragments (Acosta et al. 2001, 369).

Change of P. caesia colony size

The present study showed that there was no effect from growth and mortality rates in change of *P. caesia* colony size. Colony sizes marked in the pilot study were reported to show the difference in change of colony sizes. The results showed that large colony changed to small colonies than large colonies in the same station, such as fission in medium and large colonies of *P. caesia* on Australia's Great Barrier Reef (Tanner 1999, 390; Tanner 2000,

514). Why large colonies try to shrinkage to small colonies, Acosta (2001, 976) explained that *Palythoa* was likely to exhibit fragmentation, reducing potentially high levels of colony mortality and costs associated with reproduction. This point may answer the question of why large colonies change in small colonies.

Reproduction

P. caesia has two reproductive modes, hermaphrodite and sterile with spawning (Ryland and Babcock 1991, 117-123; Soong et al. 1999, 338). P. caesia produced gonads throughout the year and highly percent of fertilized polyps per colony were observed in cool month periods. Reduce in percent of fertilized polyps per colony were observed during rainy season, may be explained by salinity reduction in this season, probably making gametogenesis difficult (Cooke 1976, 283; Fadlallah et al. 1984, 87-88). Recently, Sphenopus marsupialis is known as brooded (Soong et al. 1999, 340). This agrees with previously reports of zoanthids (Kimura et al. 1972, 621; Yamazato et al. 1973, 275; Cooke 1976, 281; Fadlallah et al. 1984, 80; Ryland and Babcock 1991, 229). Gonads of all stages of P. caesia were found throughout the year. Some previous studies reported on reproduction modes and reproduction cycles of Zoanthids that are showed in table 9 (Kimura et al. 1972, 621; Yamazato et al. 1973, 275; Cooke 1976, 281; Fadlallah et al 1984,

Sex allocation was examined by Yamazato et al. (1973, 275-283) Fertile *Palythoa tuberculosa* polyps were mostly male but fertile *Zoanthus pulchellus* polyps studied by Karlson (1981, 699-704) were mostly hermaphrodite as in *Protopalythoa* sp. (Ryland and Babcock 1991, 117-123) and *Palythoa caesia* (this study).

Table 10 Mode of reproduction, sex, oocyte and sperm diameter and reproduction cycle in the different zoanthid studied to date. (F: Female, M: Male, H: Hermaphrodite, St: Sterile, S: Spawner, B: Broadcaster -: Indicated that information was not supplied by the authors and *: Indicated that information was derived occasional

measurements.)

			Maximum			
Species	Sex	Mode	Oocytes Diameter	Maximum Sperm	Keproductio	Source
			(µ m)	Diameter (µm)	n cycle	
					August –	Kimura and
Palythoa tuberculosa F M H St S	H N	V.	300 - 500	170- 300	November,	Yamazato. 1972;
	1) 141, 11, U	2		170-200	October -	Yamazato. 1973
					December	
Palythoa vestitus	H M H	v	080 - 36	000	Throughout	
Zoanthus pacificus	1, 171, 11, 51	,	7 - 200	065 - 06	the year	C00ke. 19/0

Table 10 (Continued)

Species	Sex	Mode	Maximum Oocytes	Maximum Sperm	Reproductio	Source
			Diameter (µm)	Diameter (µm)	n cycle	
Zoanthus solanderi	F, H, St	W	134.8 – 239.5	1	F	1 1 1 1
Zoanthus sociatus	H, St	S	64.4 - 201.0	ı	i nrougnout	radialian et al.
Palythoa caribaeorum	F, M, H, St	S	134 - 429	ı	tne year	1984
Protonalythoa sn	ъ Н	v	1 5 8 8 mm	350	July -	Ryland and
de nom (indoor	10 (11 (1	2	0.0	007	December	Babcock. 1990
Sphenomis marsunialis	F H M	α V	180 - 200	,	Throughout	Soong et al. 1999
camidus mu sudouodo	1, 11, 171		007 001	1	the year	
Palythoa caesia	j H	v	346 - 408	173 - 787	Throughout	*This study
	12 (11	מ	024 047	707 - 671	the year	

Population density and Distribution pattern

In general, zoanthids were found in shallow water (Karlson 1980, 895; Suckhnek and Green 1981, 680; Burnett et al. 1994, 154; Bastidas and Bone 1996, 544). At Khang Khao Island, P. caesia was found from 2 to 5 m. in depth and grew on four types of substrates, rock, dead coral, sand with rubber and living coral as similar to certain reports studied on distribution of zoanthids elsewhere (Fadlallah et al. 1984, 82; Burnett et al. 1997, 58; Acosta et al. 2001, 365). In addition to distribution pattern, there are several factors, such as storm damage, disturbance by sea urchin Diadema antillarum and algae can control distribution patterns of zoanthids. They are usually susceptible to storm damage that has an effect to mode of nutrition as noted in the literature review. The zoanthids well attached to occupied substrates (Karlson 1983, 128) or bare substrates (Bastides and Bone 1996, 543). Distribution of zoanthids can be as defension strategies in competition for space (Suchanek and Green.1981, 679; Bastides and Bone 1996, 550). Zoanthids had a slow growth rate when were in competition with other species by decreasing in percent cover mainly due to mortality caused by invasion of marcroalgae (Bastides and Bone 1996, 547).

The population density of *P. caesia* was reviewed in literature review elsewhere. The results from this study do support in certain points.

Competition

Zoanthid was a dominant competitor with reef sessile organisms. Studies of competition of *P. caesia* at Khang Khao Island, Chonburi Province, revealed that three groups of sessile organisms are in contact with *Palythoa*. It can overgrowth scleratinian corals, other zoanthids and sponges. Some previous reports also showed subordinate species which was overgrowth by rapid growth of Zoanthidea. Karlson (1980, 896) reported seven groups of sessile organisms showing overgrowth, *Zoanthus*, Actiniaria, Zoanthidea, sclerati-nian corals, Gorganacea, Corallimorpharia corals and Hydrocorallina. Suchanek and Green (1981, 681) reported four groups of sessile organisms showing interactions with *Palythoa caribaeorum*, scleratinian corals, Hydrozoan corals, zoanthid and sponges.

It seems clear that in a relatively short time, *Palythoa* should be able to cover even more area on the reef than it does. Only two species are know to have the ability to overgrow *Palythoa*, the encrusting gorgonian *Erythropodium caribaeoum* (Karlson 1980, 895) in Jamaica an encrusting colonial tunicate *Trididemnum solidum* (Suchanek and Green 1981, 683) in Panama. What, then, limit growth and/or abundant of *Palythoa* Suchanek and Green (1981, 683) explains by two reasons. First, the genus *Palythoa* is characterized by having encrustations in the body wall discouraging predation. Secondly, the presence of palytoxin may question of space. Although the competition strategy need more study.

Assimilated and surface sediment in P. caesia tissues

The mean assimilated sediment P. caesia tissues at Khang Khao Island was 2,844.92 g/ash/m² or 71.87% of dry tissues weight. The grain sizes in their tissues were between 20-760 μ . From the assimilated sediment investigated by Haywick and Mueller (1997, 39-46) the amount of assimilated sediment in P. caribaeorum and P. mammilosa at Florida Key, USA. Were 45% of wet tissues weight. The grain sizes were between < 63-250 μ m. The strong vary of assimilated sediment between P. caesia and Palythoa spp. tissues be able to explained by the differentce of tick mesoglea of P. caesia and high sedimentation rates at Khang Khao Island.

Surface sediment on *P. caesia* colony had a relationship with sediment-tation rate from sediment traps. This result is agreeable with Haywick and Mueller (1997, 36-46) and Acosta (2001, 119-120) The sediment defend mechanism as note in literature review. The importance of *P. caesia* in coral community changes in the Gulf of Thailand.

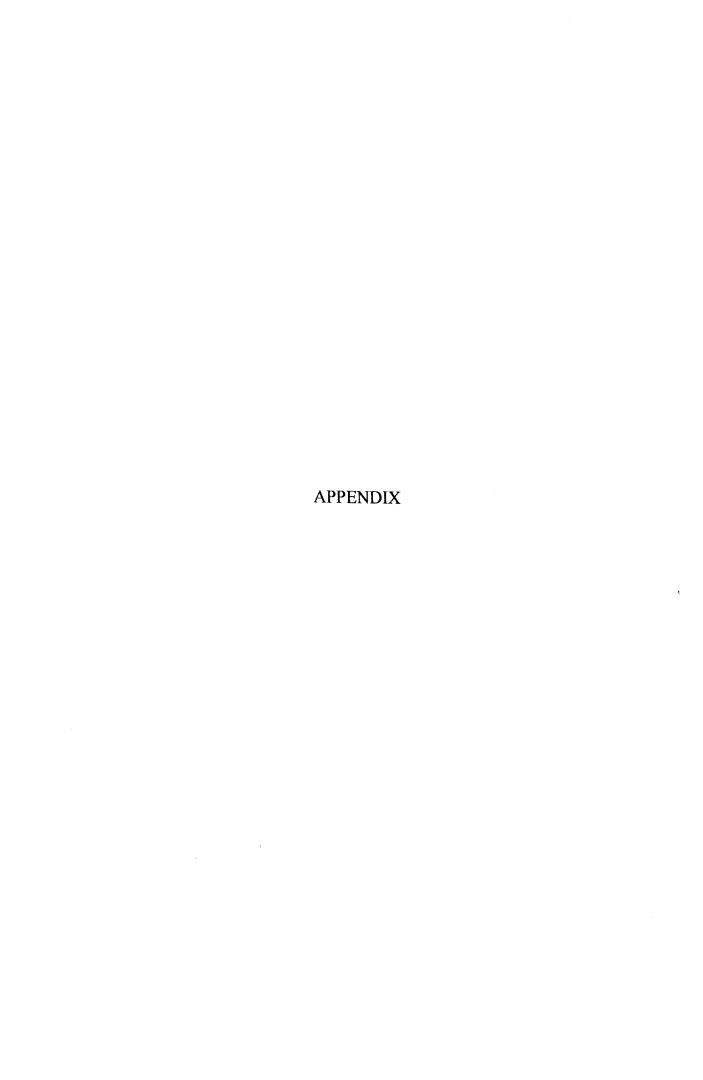


Table 11 Two factor of ANOVA on Growth and Mortality rates of P. caesia

Source of Variation	SS	df	MS	F	P-value	F crit
Rows (Month)	81.873	16	5.1171	2.257	0.006	1.7231
Columns (Station)	95.516	8	11.94	5.266	0.000010	2.0115
Error	290.21	128	2.2673			
Total	467.6	152				

Table 12 Two factor of ANOVA on 5 cm. diameter sediment traps and sediment plates

Source of						
Variation	SS	df	MS	F	P-value	F crit
Rows	22.5046		4.510010	0.020070	0.524025	2 202026
(Station)	22.5946	5	4.518919	0.829879	0.534025	2.382826
Columns	102.0124	1 1	17.54650	2 222251	0.001000	1.067546
(Traps)	193.0124	11	17.54658	3.222351	0.001923	1.967546
Error	299.49	55	5.445273			
Total	515.097	71				

Table 13 Two factor of ANOVA on 1.5 cm. diameter sediment traps and sediment plates

Source of Variation	SS	df	MS	F	P-value	F crit
Rows	220 525 6	1	220.525.6	24.00252	0.004200	7.70065
(Station)	230,535.6	1	230,535.6	34.09253	0.004289	7.70865
Columns	27.054.19	4	6 762 544	1.00022	0.4000177	6 200224
(Traps)	27,054.18	4	6,763.544	1.00022	0.499917	6.388234
Error	27,048.22	4	6,762.055			
Total	284,638	9				

Table 14 Two factor of ANOVA on surface sediment from *P. caesia* and resuspension of bottom sediment

Source of Variation	SS	df	MS	F	P-value	F crit
Rows (Sediment rates)	304,325,4 66.40	2	1,521,627, 33.20	1.002355	0.499412	19.00003
Columns (Station)	27,908,81 8,993.19	1	27,908,81 8,993.19	183.8462	0.005395	18.51276
Error	303,610,4 51.3	2	151,805,2 25.67			
Total	28,516,75 4,911	5				

Table 15 Two factor of ANOVA on 1.5 cm. diameter and 5 cm. diameter sediment trap

Source of						
Variation	SS	df	MS	F	P-value	F crit
Rows	22.5046		4.510010	0.000070	0.524025	2 202026
(Month)	22.5946	5	4.518919	0.829879	0.534025	2.382826
Columns	102 0124	1 1	17 54650	2 222251	0.001022	1.065546
(Station)	193.0124	11	17.54658	3.222351	0.001923	1.967546
Error	299.49	55	5.445273			
Total	515.097	71				

Table 16 Two factor of ANOVA on 1.5 cm. diameter sediment trap and sediment plate

Source of						
Variation	SS	df	MS	F	P-value	F crit
Rows	27.054.10		C 51 CO 5 1 1	1 00000	0.40004.7	
(Traps)	27,054.18	4	6,763.544	1.00022	0.499917	6.388234
Columns	220.525.6	1	000 505 6	24.002.52	0.004000	
(Station)	230,535.6	1	230,535.6	34.09253	0.004289	7.70865
Error	27,048.22	4	6,762.055			
Total	284638	9				

Table 17 Two factor of ANOVA on 5 cm. diameter sediment trap and sediment plate

Source of						
Variation	SS	df	MS	F	P-value	F crit
Rows	420 5202	4	105 0005	0.000=1.4		
(Traps)	429.5302	4	107.3825	0.993714	0.502365	6.388234
Columns	1 (07 20)	1	1 (07 00)	140500	0.040=04	
(Station)	1,607.206	1	1,607.206	14.87303	0.018201	7.70865
Error	432.2473	4	108.0618			
Total	2,468.984	9				

BIBLIOGRAPHY

- Acosta., A. 2001. Disease in zoanthids: dynamics in space and time. **Hydrobiologia** 460: 113-130.
- Acosta, A., P. W. Sammarco, and L. F. Duarte. 2001. Asexual reproduction in a zoanthid by fragmentation: the role of exogenous factors. **Bull.**Mar. Sci. 68:363-381
- Aerts, L. A. M., and R. W. M. van Soest. 1997. Quantification of sponge/coral interactions in a physically stressed reef community, NE Colombia.Mar Ecol. Prog. Ser. 148: 125-134.
- Allen, G. R., and R. Steen. 1999. Indo-Pacific Coral Reef Field Guide. 5d ed. 55-64. Singapore: Calendar Print.
- Amano, K., K. Sato, M. Hori, H. Ozaki, and H. Karaki. 1997. Palytoxin-induced increase in endothelial Ca²⁺ concentration in the rabbit aortic valve. **Archives of Pharmacology** 355: 751-758.
- Anthony, K. R. N., and K E. Fabricius. 2000. Shifting roles of heterotrophy And autotrophy in coral energetic under varying turbidity. J. Exp. Mar. Bio. Ecol. 252: 221-253.
- Babcock, R. C., and J. S. Ryland. 1990. Larval development of a tropical zoanthid (*Protopalythoa* sp.). **Invert. Reprod. Devel**. 17: 229-236.
- Babcock, R. C., and P. Davies. 1991. Effects of sedimentation on settlement of *Acropora millepora*. **Coral reef** 9: 205-208.

- Barnes, R. S. K., P. Calow, P. J. W. Olive, and D. W. Golding. 1988. Phylum Cnidaria (hydras, jellyfish, anemones, corals). In **The Invertebrates**, 55-70. London: Blackwell Scientific Press.
- Bastidas, C., and D. Bone. 1996. Competitive strategies between *Palythoa* caribaeorum and *Zoanthus sociatus* (Cnidaria: Anthozoa) at a reef flat environment in Venezuela. **Bull. Mar. Sci.** 59: 543-555.
- Berntson., E. A., S. C. France, and L. S. Mullineaux. 1999. Phylogenetic relationships within the class Anthozoa (Phylum Cnidaria) based on nuclear 18S rDNA sequences. **Mole. Phyl. Evol.** 13: 417-433.
- Borneman, E. 1998. Zoanthids Coral? Anemone? Both? Neither?.

 Aquarium. Net January. [Online]. Available URL: www. aqarium.net.
- Borneman., E.H., and J. Lowrie. 2001. Advances in captive husbandry and propagation: an easily utilized reef replenishment means from the private sector?. **Bull. Mar. Sci.** 69: 897-913.
- Buchanan, J. B. 1984. Sediment analysis. In: Methods For the Study of Marine Benthos, ed. N. A. Holme, 41-65. London: Blackwell Scientific Press.
- Burnett, W. J., J. A. H. Benzia, J. A. Beardmore, and J. S. Ryland. 1994. High genetic variability and patchiness in a common Great Barrier Reef zoanthid (*Palythoa caesia*). **Marine Biology** 121: 153-160.
- _____. 1995. Patterns of genetic subdivision in populations of a clonal cnidarian, *Zoanthus coppingeri*, from the Great Barrier Reef. Marine Biology 122: 665-673

- Burnett, W. J., J. A. H. Benzia, J. A. Beardmore, and J. S. Ryland. 1997.

 Zoanthids (Anthozoa, Hexacorallia) form the Great Barrier Reef and

 Torres Striat Australia: Systemaics, Evolution and key to species.

 Coral Reefs 16: 35-68.
- Campbell, R. D. 1974. Cnidaria. In Reproduction of Marine
 Invertebrates. Vol. 1, ed. A. C. Giese, 133-199. New York: Acaemic Press.
- Carte, B. K. 1996. Biomedical potential of marine natural products. **Bio. Science**. 46: 271-287.
- Cooke, W. J. 1976. Reproduction, growth and some tolerances of
 Zoanthus pacificus and Palythoa vestitus in Kaneohe Bay Hawaii.

 In Coelenterate Ecology and Behavior, ed. G. O. Mackie, 281-288.

 New York: Plenum Press.
- Chulabhorn Research Institute and Thai Royal Navy. 1998. Guide of coral reef fauna and flora of Surin and Similan Islands. n.p.
- Eckert, R. 1988. The restung potential. In Animal physiology mechanism and Adaptation. 3d ed. 149-173 New York: Freeman Press.
- English, S., C. Wilkinson, and V. Baker. 1997. Survey Manual for Tropical Marine Resources. 2d ed. Australian: Institute of Marine Science, Townsville.
- Fadlallah, Y. H, R. H. Karlson, and K. P. Sebens. 1984. A comparative study of sexual reproduction in three species of Panamanian zoanthids (Coeleterata: Anthozoa). **Bull. Mar. Sci.** 35: 50-89.

- Gleibs, S., D. Mebs, and B. Werding. 1995. Studies on the origin and distribution of palytoxin in a caribbean coral reef.Toxicon 33: 1531-1537.
- Gleibs, S., and D. Mebs. 1999. Distribution and sequestration of palytoxin in coral reef animals. **Toxicon** 37: 1521-1527.
- Hadfield, M. G., E. J. Carpizo-Ituarte, K. del Carmen, and B. T. Nedved. 2001. Metamorphic competence, a major adaptive convergence in marine Invertebrate larvae. Amer. Zool. 41: 1123-1131.
- Haywick, D. W., and E. M. Mueller. 1997. Sediment retention in encrusting *Palythoa* spp. a biological twist to geological process.Coral Reefs 16: 36-46.
- Hill, M. S. 1998. Sponges harbor genetically identical populations of the zoanthid *Parazoanthus parasiticus*. **Bull. Mar. Sci.** 63: 513-521
- Karlson, R. H. 1980. Alternative competitive strategies in a periodically disturbed habital. **Bull. Mar. Sci.** 30: 894-900.
- Bay, Jamaica. In **Proc.4**th. **Int. Coral Reef Symp**. Vol. 2, 699-704.

 Manila: University of the Philippines.
- _____. 1983. Disturbance and monopolization of spatial resource by Zoanthus socitus (Coelenterata, Antrozoa). Bull. Mar. Sci. 33: 118-131.
- _____. 1988a. Growth and survivorship of clonal fragments in Zoanthus solanderi Lesueur. J. Exp. Mar. Bio. Ecol. 123: 31-39.

- Karlson, R. H. 1988b. Size-dependent growth in two zoanthid species: a contrast in clonal strategies. **Ecology** 69: 1219-1232.
- Kimura, S., Y. Hashimoto, and K. Yamazato. 1972. Toxicity of the zoanthid *Palythoa tuberculosa*. **Toxicon** 10: 611-617.
- Koehl, M. A. R. 1977. Water flow and the morphology of zoanthid colonies.

 In **Proc 3rd Int Coral Reef Symp.** Vol. 1, 437-444. Manila: University of Miami.
- Leech, C. A., and J. F. Habener. 1998. A role for Ca²⁺-sensitive nonselective cation channels in regulating the membrane potential of pancreatic beta-cells. **American diabetes Association** 47: 1066-1075.
- Martindale, M. Q., and J. Q. Henry. 1998. The development of radial and biradial symmetry: the evolution of bilaterality.

 Amer. Zoo. 38: 672-684.
- Mebs, D. 1998. Occurrence and sequestration of toxins in food chains.

 Toxicon. 41-43
- Moore, R. E., and P. J. Scheuer. 1971. Palytoxin: a new marine toxin from a Coelenterate. **Science** 172: 495-498.
- Rogers, C. S. 1983. Sublethal and lethal effects of sediments applied to Common Caribbean Reef Corals in the field. Marine Pollution

 Bulletin 14: 378-382
- Rhodes, L., N. Towers, R. Munday, and J. Adamson. 2002. Uptake of palytoxin-like compound by shellfish fed *Ostreopsis siamensis* (Dinophycae). New Zealand Journal of Marine and Freshwater Research 36: 631-636.

- Roy, K. J., and S. V. Smith. 1971. Sedimentation and coral reef development in turbid water: Fanning Lagoon. **Pacific Science** 25: 234-248.
- Ruengsawang, N. 1999. Bioerosion by sea urchin, *Diadema setosum*, in a coral community at Khang Khao Island, Inner Gulf of Thailand.M. Sc. Thesis, Ramkhamhaeng University.
- Ryland, J. S., and R. C. Babcook. 1991. Annual cycle of gametogenesis and spawning in a tropical zoanthid, *Protopalythoa* sp. **Hydrobiologia** 16, 217: 117-123.
- Scheiner-Bobis, G., D. M. Z. Heringdorf, M. Christ, and E. Habermann. 1994.

 Palytoxin Induces K⁺ Efflux form yeast cells expressing the

 mammalian Sodium pump. **Molecular Pharmacology** 45: 1132-1136
- Sebens, K. P. 1977. Autotrophic and heterotrophic nutrition of coral reef zoanthids. In **Proc 3rd Int Coral Reef Symp**. Vol. 1, 397-408. Manila: University of Miami.
- Soong, K, Y. S. Shain, and C. P. Chen. 1999. Morphological and life history divergence of the zoanthis, *Sphenopus marsupialis* off the Taiwanese Coast. **Zoological studies** 38(3): 333-343.
- Suckhnek, T. H., and D. J. Green. 1981. Interspecific competition between Palythoa caribaeorum and other sessile invertebrates on St. Croix Reefs, U.S. Virgin Island. In **Prog.4**th. Int. Coral Reef Symp. Vol. 2, 679-684. Manila: University of Philippines.

- Sudara, S., A. Sanitwong, T. Yeemin, R. Moordee, S. Panutrakune, P. Suthanaluk, and S. Natekanjanalarp. 1991. Study of the impact of sediment on growth of the coral *Porites lutea* in the Gulf of Thailand. In **Proceedings of the Regional Symposium Living Resources in Coastal Areas**. ed. R. M. de la Paz, 107-112. Manila: University of the Philippines.
- Suthanarak, P. 1983. Studies on effects of sedimentation on growth rates of some scleractinian corals from Khang Khao island, Chonburi

 Province. Special Problem Report. Bankok: Department of Marine Science, Chulalongkorn University.
- Tanner, J. E. 1997. The effects of density on the zoanthid *Palythoa casia*. J. Ani. Ecol. 66: 793-810.
- . 1999. Density-dependent population dynamics in clonal organisms: a modelling approach. J. Ani. Ecol. 68: 390-399.
 . 2000. Stochastic density dependence in population size of a benthic clonal invertebrate: the regulating role of fission. Oecologia
- Tosteson, M. T., D. R. L. Scriven, A. K. Bharadwaj, Y. Kishi, and D. C. Tosteson. 1995. Interaction of palytoxin with red cell: structure-function studies. **Toxicon** 33(6): 799-807

122: 514-520.

Yamazato, K., F. Yoshimoto, and N. Yoshihara. 1973. Reproductive cycle in a zoanthid *Palythoa tuberculosa* Esper. **Publ. Seto. Mar. Biol. Lab**. 20: 275-283.

- Yamazato, K., F. Yoshimoto, and N. Yoshihara. 1987. Effects of deposition and suspension of inorganic particulate matter on the reef building corals in Okinawa, Japan. **Galaxea** 6: 289-309.
- Wessling, I., A. J. Uychiaoco, P. M. Aliño, T. Aurin, and J. E. Vermaat. 1999.

 Damage and recovery of four Philippine corals from short term sediment burial. Mar. Eco. Prog. Ser. 176: 11-15
- Wu, C. H., L. A. Vasilets, K. Takedad, M. Kawamurad, and W. Schwarzb.

 2003. Function role of the N-terminus Na⁺,K⁺-ATPase α-subunit as an Inactivation gate of palytoxin-induced chanenel. **Biochim. Biophys.**Acta. 1609: 55-62

VITA

Miss. Napalai Juntaruk was born on May, 22 1977 in Bangkok Province. She finished her high school education from Nawamintrachuthid Bangkok School in 1995. She graduated with Bachelor Degree of Education (Biology) from Division of Biology Rajabhat Institute Phanakhon in 1999. After that she studied for a Master Degree in Biology at the Ramkamhang University. Her research outputs are presented in the national conferences are shown as follow:

- Juntaruk, N., J. Buaruang and T. Yeemin. 2000. Sediment retention in a Zoanthid, *Palythoa* sp. from The Inner Gulf of Thailand. Poster presented at 26th Congress on Science and Technology of Thailand. Bangkok, Thailand, 18-20 October.
- Juntaruk, N. 2001. Ecology of *Palythoa* sp. (Anthozoa: Zoanthidea) in
 Turbid environment. Poster presented at the 5th Congress on
 Biodiversity Research and Training Program. Udonthani, Thailand, 8-10 October.
- ______. 2002. Ecology of *Palythoa* sp. (Anthozoa : Zoanthidea) in

 Turbid environment.. Nakorn-srithammarat, Poster presented at the 6th

 Congress on Biodiversity Research and Training Program. Nakon

 Sithammarat, Thailand, 9-11 October.

Juntaruk, N. 2003. Ecology of *Palythoa ceasia* (Anthozoa: Zoanthidea) in Turbid Environment Poster presented at the 7th Congress on Biodiversity Research and Training Program. Chiang Mai, Thailand, 13-16 October.